

**EVALUATING THE LEGACY OF SEVERE WILDFIRE ON AQUATIC
CARBON DYNAMICS AND MICROBIOMES IN MONTANE
WATERSHEDS**

A Thesis

Presented to

The Faculty of the Environmental Program

The Colorado College

In Partial Fulfillment of the Requirements for the Degree

Bachelor of Arts in Environmental Science

By

Michelle A. Wolford

May 2021



Dr. Rebecca T. Barnes
Associate Professor of Environmental Science, Colorado College



Dr. Michael J. Wilkins
Associate Professor of Soil and Crop Science, Colorado State University

TABLE OF CONTENTS

ABSTRACT	1
ACKNOWLEDGEMENTS	2
INTRODUCTION	4
Effects of Wildfire on Aquatic Systems.....	4
Post-fire Shifts in Microbial Community Composition	8
Research Questions	9
METHODS	11
Site Description	11
Sampling and Measurements.....	12
<i>Stream Water Collection</i>	12
<i>Solute and DOC Analysis</i>	13
<i>Isotopic Analysis</i>	14
<i>Metabolism Measurements</i>	14
Microbial Community Characterization.....	16
Data Analysis	16
RESULTS	18
Watershed Characterization	18
Stream Solutes.....	18
<i>Dissolved Organic Matter</i>	18
<i>$\delta^{13}\text{C}$-DOC</i>	20
<i>Inorganic Carbon</i>	21
<i>$\delta^{13}\text{C}$-DIC and $\delta^{13}\text{C}$-CO₂</i>	21
<i>Nitrogen</i>	22
<i>$\delta^{18}\text{O}$-H₂O</i>	23
Stream Metabolism	23
<i>In Situ Incubations</i>	23
<i>Dissolved C Bioavailability</i>	23
<i>Modeled Daily Metabolism</i>	24
Microbial Communities.....	25
<i>Diversity Metrics</i>	25
<i>Differences in Relative Abundance</i>	26
Changes to C Export and Processing	27
Shifts in the Stream Microbiome.....	30
CONCLUSION	34
TABLES AND FIGURES	35
REFERENCES	52

ABSTRACT

As the climate continues to warm, wildfires are expected to increase in frequency and severity, particularly across the western United States, likely impacting the global carbon (C) balance. Fire influences C stocks via shifts in vegetation, hydrologic flow paths, and microbial communities. These disturbances alter C partitioning across landscapes, connectivity between hillslopes and streams, and microbial function. However, potential linkages between disturbance, C cycling, and terrestrial and aquatic microbiomes require further investigation. This study explored the long-term legacy of wildfire on watershed C dynamics and aquatic microbial communities, while evaluating how changes to the terrestrial landscape are reflected in stream systems. During the summer of 2019, dissolved inorganic and organic carbon pools were measured along with estimates of stream metabolism and characterization of aquatic microbiomes for three streams draining catchments that burned during the 2002 Colorado wildfire season and two streams unaffected by fire. Compared to unburned watersheds, aquatic C exports and microbiomes were characteristically different in burned watersheds; specifically, C stocks were less aromatic and less bioavailable, which could exert controls on microbial communities. Although stable isotope analyses revealed that all streams were dominated by allochthonous sources, trends in dissolved C pool composition in streams draining unburned landscapes suggest shifting flow paths and/or differences in in-situ sources and transformation between watersheds. Overall, wildfire appears to create a feedback loop between C quality, processing, and microbiomes due to changes in vegetation and hydrology in the terrestrial environment as well as differences in microbiomes across the landscape. These alterations could affect recovery dynamics in burn scars and imply that wildfires in warmer, drier climates could result in steady state shifts.

ACKNOWLEDGEMENTS

First and foremost, I would like to thank my advisor and mentor, Dr. Rebecca Barnes. In countless ways, Becca has made me the scientist—and human—that I am today. She brought me onto this project the summer after my sophomore year at Colorado College, with very little experience in the field, and I am eternally grateful for her guidance and encouragement that followed. Simply reminding me of how much I learned since the beginning of this process means so much to me, let alone the countless hours spent on Zoom talking all things water, carbon, and even imposter syndrome. From baking our lab group bread on the weekends to leaving us notes and geoscience-related stickers, my thesis would not have made it this far without Becca's endless support. She taught me that I belong in this field. I am immensely thankful for Becca's mentoring and kindness the last three years.

I would also like to offer my appreciation to my collaborators at Colorado State University, Dr. Michael Wilkins and Amelia Nelson. Mike and Amelia helped characterize the microbial communities for this project and have shared valuable knowledge with me about microbial ecology. I am thankful for Mike's words of encouragement and detailed feedback on my writing. I am also extremely grateful for Amelia's assistance and patience. Whether it was processing our microbial data, teaching me multivariate statistics, troubleshooting R code, or thinking about paper organization, Amelia has been my biggest cheerleader, even if only over Slack messages.

Additionally, my gratitude extends to many members of the Environmental Program at Colorado College. Cosette Turvold and Oliver Dunn created a welcoming and inclusive environment in the lab this year, and I am thankful for all their advice about figures and writing. It was a blessing to have such great peers by my side. I want to specifically thank Coco for the

conversations about our futures as scientists and offering many thesis writing-themed jokes; she always managed to make me smile even when the work felt overwhelming. Additionally, I am appreciative of the academic support and great friendship from my past lab colleagues, Carly Bonwell and Cheristy Jones. Their enthusiasm in exhausting field conditions and commitment during long days in the lab were integral to this project. Together, Carly and Cheristy showed me how rewarding research can be, and their guidance (including mailing me a handwritten list of 15 “Thesis Tips”) has shaped me in many ways, and I am indebted to Cherry for our many phone calls about data, grad school, and more. I could not have done this without these four lovely science friends. I also thank Darren Ceckanowicz for his assistance in the laboratory and providing technical support during our field season in 2019.

Lastly, I would like to thank my friends and family who cared for me during such unprecedented times, checked in on me and my research, and celebrated little wins along the way—like jumping for joy when my code finally ran or hugging me when I finished my first draft. These wonderful humans always stood by my side even if they had never heard of pyrogenic carbon or Actinobacteria before. This would not have been possible without every person who helped along the way.

INTRODUCTION

Effects of Wildfire on Aquatic Systems

Wildfire represents a dominant disturbance that influences landscapes on a global scale (Bowman *et al.*, 2009), and, as the climate continues to warm, fires are expected to increase in both frequency and severity (IPCC, 2018). The negative impacts of increasing wildfire are numerous and extend to many groups of people and ecosystem services. Air pollution from smoke (Reid *et al.*, 2016), habitat loss (Morton *et al.*, 2003), changes to water quality (Chow *et al.*, 2019), and increased hazard for subsequent natural disasters (Moody & Martin, 2001) during and after fire pose great risks for many populations in the face of climate change (IPCC, 2018). Such disturbances can also reduce biodiversity (Abella & Fornwalt, 2015) and alter long-term nutrient storage (McLauchlan *et al.*, 2014), endangering the longevity and health of ecosystems that offer needed services (IPCC, 2018). Additionally, fire destroys forests that naturally sequester significant amounts of carbon (C) from the atmosphere, which threatens the global C balance and augments the greenhouse gas effect (IPCC, 2018).

As drier and warmer conditions persist in the western United States (U.S), fire return intervals will likely shorten, yielding little recovery time for these forested environments (Enright *et al.*, 2015). Since the 1980s, fire season in this region has become longer, and fires have increased in numbers and duration (Westerling *et al.*, 2006; Abatzoglou & Williams, 2016). Montane forests in the northern Rocky Mountains are at particular risk for fire as spring and summer temperatures rise, spring snowmelt occurs earlier, and drought-related impacts increase (Westerling *et al.*, 2006; Enright *et al.*, 2015; Abatzoglou & Williams, 2016; Westerling *et al.*, 2016). For example, the Hayman Fire of 2002, which prior to 2020 was the largest wildfire in recorded Colorado history, burned 560 km² of land southwest of Denver, CO.

However, in the summer and fall of 2020, the Cameron Peak Fire, East Troublesome Fire, and Pine Gulch Fire all grew much larger than the Hayman Fire, burning over 2,100 km² in northern and western parts of the state (Johnson, 2020; Kirk, 2020; USFS, 2020). These fires directly harmed local Colorado populations, causing over \$195 million in damage, unsafe air quality levels, and the relocation of many communities (Miller, 2020); still, the lasting socio-economic costs of the 2020 fire season will likely be much more. The short- and long-term effects, some of which are enumerated here, only emphasize the imminent dangers posed by increasing wildfires in the west.

Understanding the consequences of changing fire regimes in the western U.S. on C partitioning is particularly important as current research suggests that forests in this region constitute 20 to 40% of all carbon sequestration in the country (Pacala *et al.*, 2001; Huber *et al.*, 2006; Westerling *et al.*, 2006). However, persistent changes to vegetation and soil organic matter (SOM) following fire limit the ability of these systems to effectively sequester and store C (Buma *et al.*, 2014; Fornwalt *et al.*, 2016). In essence, Ponderosa pine (*Pinus ponderosa*) forests, like those studied here, historically function as carbon sinks, but new fire activity and slow rates of recovery may shift these systems toward serving as carbon sources. Losses of these C sinks will only exacerbate the negative repercussions of climate change. These landscapes will not be able to efficiently remove C emitted by anthropogenic sources (e.g., burning fossil fuels) from the atmosphere, resulting in significant management implications: if wildfire disturbs a majority of global forests so they no longer effectively store C, the global C balance may be altered for centuries.

By modifying terrestrial landscapes, fire also changes nutrient export (Knicker, 2007). Fire yields greater runoff and higher stream discharges in response to changes in vegetation

(Buma *et al.*, 2013; Ebel & Moody, 2013) and/or soil structure (González-Pérez *et al.*, 2004; Rhoades *et al.*, 2011), which can greatly alter hydrology by changing flow paths and decreasing rates of evapotranspiration (Hallema *et al.*, 2017). Such disturbances allow for the formation of preferential flow paths and overall decreased soil water holding capacity (Gilbertson 2018; Barnes & Gilbertson, 2018, unpublished data). Lack of vegetation and thus belowground biomass also reduce demand for nutrients within the watershed, increasing transport to stream systems (Certini *et al.*, 2005). Given that nutrient and C mobilization is strongly linked to hydrologic transport through the landscape (Xu & Saiers, 2010) and the magnitude of hydrological responses to fire (Moody & Martin, 2001), these differences likely control C export—and storage—as well.

Allochthonous C sources (e.g., C inputs from the watershed) dominate small headwater streams (Vannote *et al.*, 1980; Palmer *et al.*, 2001; Hotchkiss *et al.*, 2018), like those studied here. In many systems, concentration-discharge relationships suggest a transport limitation or a chemostatic response of dissolved organic carbon (DOC) to hydrologic fluctuations (i.e., mobilization is proportional to changes in watershed discharge) (Godsey *et al.*, 2009; Godsey *et al.*, 2019). Given that DOC concentrations often respond positively to flow events (Striegl *et al.*, 2005; Raymond & Saiers, 2010; Dawson *et al.*, 2008), hydrology serves as a significant driver for headwater stream DOC fluctuations (Guarch-Ribot *et al.*, 2016).

The hydrologic connectivity of hillslopes and streams links terrestrial and aquatic environments, and the amount of C held within vegetation and soils (i.e., terrestrial C sequestration) influences lateral C flux to streams. Prior to a decade ago, the transport of C to aquatic environments was often disregarded in net ecosystem C balance calculations (e.g., Foley *et al.*, 1996; Cramer *et al.*, 2001; Prentice *et al.*, 2001). Rivers were thought of as “pipes” to

transport C (Cole *et al.*, 2007; Hotchkiss *et al.*, 2015), as DOC accounts for most of the total annual export from watersheds (Mei *et al.*, 2012) and the majority fate of C in lentic systems is export (Hotchkiss *et al.*, 2018). Yet, lateral flux to inland waters affects C storage, processing, and transport (Cole *et al.*, 2007; Butman *et al.*, 2016). For example, freshwater C fluxes—C exported downstream or emitted to the atmosphere—provide 106 Tg C per year in the contiguous U.S. (Butman *et al.*, 2016), which represents a much larger contribution than previously considered in C budgets. When leaching and export of C is ignored in the equation, terrestrial net ecosystem production (NEP) and CO₂ uptake and storage by the landscape can be overestimated (Hotchkiss *et al.*, 2015); export could represent up to 27% of NEP estimations (Butman *et al.*, 2016). Given how fire changes terrestrial C stocks and the importance of inland waters to global C cycling, it is necessary to understand how fire affects lateral C export to streams draining burned watersheds and the subsequent fate of this C.

Although terrestrially derived sources dominate small streams, in-stream processes, such as microbial respiration, also influence the C pool (Hotchkiss *et al.*, 2015), and fire may shift autochthonous processing of C by altering in-situ conditions (Rhodes *et al.*, 2011; Murphey *et al.*, 2018). Estimates of stream metabolism—ecosystem respiration (ER), gross primary production (GPP), and net ecosystem production (NEP = GPP - ER)—are effective proxies for measuring stream productivity and processes (Betts & Jones, 2009; Hotchkiss & Hall, 2014). For example, ecosystem respiration has been shown to be highest in streams draining burned watersheds compared to those draining unburned watersheds (Betts & Jones, 2009). Moreover, differences in photosynthetically active radiation (PAR), primary productivity (Mulholland *et al.*, 2001), and discharge (Finlay, 2003), all of which may be impacted by severe fire, can change ecosystem respiration rates and other in-stream processes.

Post-fire Shifts in Microbial Community Composition

Fire transforms the size, activity, and composition of microbial biomass in the terrestrial environment (Hart *et al.*, 2005) and both directly and indirectly affects the stream microbiome. Shifts in microbial community composition immediately after fire are often due to heat-induced mortality during heat transfer that occurs with the combustion of organic matter (OM) (Hart *et al.*, 2005; Pressler *et al.*, 2019). Altered environmental conditions (e.g., SOM character, etc.) following fire may also lead to differences in communities (Knicker, 2007). Given that the composition of stream microbiomes may be influenced by transport of terrestrial microorganisms from soil and upslope communities into fluvial systems (Crump *et al.*, 2003; Crump *et al.*, 2012), documented shifts in the terrestrial environment may be reflected in the aquatic microbiome (Leibold *et al.*, 2004; Vellend, 2010).

Still, little is known regarding the influence of wildfire on both terrestrial and aquatic microbial communities (Pressler *et al.*, 2019). Most bacterial phyla are not immediately resilient to disturbance regardless of taxonomic complexity or severity of disturbance events (Allison & Martiny, 2008). Thus, the environmental conditions that persist following fire (e.g., elevated pH, altered vegetation, changes in nutrient availability, etc.) likely influence microbial community composition (Vellend, 2010). In general, fire is expected to reduce microbial biomass, abundance, and diversity (Pressler *et al.*, 2019), but much of this research rests on the assumption that stream and soil communities respond similarly to fire. Thus, the response of terrestrial and aquatic microbiomes to disturbance needs to be further studied (Langenheder & Lindström, 2019).

Changes in microbiome composition and activity due to disturbance may directly affect ecosystem processes, such as respiration, given the reliance of heterotrophic microbial

community members on DOC (Kaplan *et al.*, 2008). As such, surveys of bacterial diversity in freshwater have identified many populations that are involved in the degradation of OM and other processes related to C cycling, including members of the Bacteroidetes, Cyanobacteria, Patescibacteria, Actinobacteria, and Verrucomicrobia phyla (Glöckner *et al.*, 2000; Zwart *et al.*, 2002; Eiler *et al.*, 2002). Overall, it is well understood that biogeochemical (e.g., DOM supply, Judd *et al.*, 2006) and environmental factors (e.g., pH, Fierer *et al.*, 2007) influence these aquatic microbial communities, and differences in community attributes (e.g., diversity) can govern ecosystem processes (Langenheder *et al.*, 2010; Frossard *et al.*, 2012).

The composition and activity of microbial assemblages is often linked to C quality and/or quantity (Bernhardt & Likens, 2002; Crump *et al.*, 2003; Eiler *et al.*, 2003; Judd *et al.*, 2006; Strickland *et al.*, 2009; Zeglin, 2015). Different physical and nutritional habitats in the water column may select for different microbial assemblages (Zeglin, 2015; Stelzer *et al.*, 2003) and microbial functions (Bernhardt & Likens, 2002). Organic matter lability and quality can also regulate communities, with the addition of labile C stimulating bacterial production and community shifts (Crump *et al.*, 2003; Judd *et al.*, 2006; Crump *et al.*, 2012). Since disturbances transform C transported to streams, fire likely impacts stream microbial assemblages indirectly.

Research Questions

While many studies address how wildfire alters different watershed characteristics, such as flow paths (Moody & Martin, 2001; Ebel & Moody, 2013; Hallema *et al.*, 2017), nutrient cycling (Smith *et al.*, 2011; Santos *et al.*, 2016; Rhoades *et al.*, 2018), and microbial community composition (Hart *et al.*, 2005; Knicker, 2007; Pressler *et al.*, 2019), further investigation is still required to synthesize the positive feedback loops between disturbance, C cycling, and microbiomes. Even research that considers how wildfire influences C sequestration and

partitioning across the landscape (Buma *et al.*, 2014) often overlooks linkages between hillslopes and streams. And, although some studies address differences in the terrestrial microbiome, few studies have been conducted surrounding aquatic microbial community structure and function following fire. Therefore, additional work needs to be done to understand the long-term legacy of fire on overall ecosystem function.

The goal of this study was to understand how C cycling and microbial community structure differ post-fire and to begin a discussion on the ways fire collectively alters terrestrial and aquatic C stocks and microbial communities. Building off the work of Jones (2020) and Bonwell (2020), who looked at the legacy of fire on carbon dynamics and export across terrestrial and aquatic environments respectively, I investigated how shifts in C cycling regulate microbial assemblages in three streams draining landscapes burned during the Hayman Fire, CO as compared to two streams draining unburned landscapes. Research questions included: (1) How does fire change C quality and export in aquatic systems? (2) Are C sources different between burned and unburned watersheds? (3) In what manner is the fate of C in these landscapes determined by fire? (4) Does fire shift the composition of stream microbiomes? (5) How do changes in biogeochemical factors influence stream microbial assemblages? (6) Do shifts in microbial community structure predict shifts in ecosystem functioning (e.g., ecosystem respiration)?

METHODS

Site Description

The sites for this study are in and around Deckers, CO, USA in the Colorado Front Range (Table 1; Fig. 1). This region receives an annual average of 60 cm of precipitation and has an average temperature of 9.4°C (Stevens, 2013). Pikes Peak batholith underlies the majority of this area (Bryant *et al.*, 1981), and soils are weakly developed, excessively drained, coarse sandy loams (Stevens, 2013). Forests are in the lower montane zone (1980–2750 m elevation) and are dominated by Ponderosa pine (*Pinus ponderosa*) and Douglas fir (*Psuedotsuga menziesii*) with understory plants including forbs, shrubs, and graminoids (Fornwalt *et al.*, 2016).

In May 2002, the Schoonover Fire burned 16 km², and in June 2002, the Hayman Fire burned over 560 km² of the South Platte River drainage; both fires were stand-replacing. Seventeen years post-fire, watersheds in the burn scars experienced little recovery due to slow rates of tree colonization and forest regrowth and were characterized by grasses and forbs (Rhoades *et al.*, 2011; Fornwalt & Kaufmann, 2014). Unburned watersheds generally exhibited greater tree cover (Jones, 2020) and resembled the pre-fire conditions of burned counterparts, containing more live trees, woody plants, conifers, and litter cover (Fornwalt & Kaufmann, 2014). However, vegetation differed noticeably between the two unburned watersheds, Sugar and Painted Rocks—see Jones (2020) for vegetation characterization. Specifically, Painted Rocks ($12.65 \pm 0.03\%$) had significantly more grass cover than Sugar ($4.2 \pm 0.02\%$) ($p = 0.019$). Therefore, watershed land cover (i.e., vegetation) and site location possibly affected results in manners not quantified in the study.

Five first- or second-order streams were studied, each draining an independent catchment (Fig. 1). Three streams—Cabin, Pine, and Fourmile—drain catchments that burned in the 2002

fires, and watershed sizes span from 11.1 to 24.9 km², with the total area burned varying across the three catchments. Overall burn extent ranges from 28% in Cabin, 54% in Pine, and 82% in Fourmile (Table 1). Two streams—Sugar and Painted Rocks—drain landscapes that have not burned in over 100 years but are geographically close to the burned sites to serve as reference systems. Unburned watershed areas encompass that of the burned watersheds, ranging from 4.5 km² in Painted Rocks to 36.2 km² in Sugar.

Sampling and Measurements

Stream Water Collection

During the summer of 2019 (May 31-July 30), the five streams were sampled. Stream chemistry as well as velocity, depth, and width were measured weekly. Velocity was determined via one of two methods: (1) using a Swiffer Model 3000 Current Velocity and Stream Discharge Indicator in streams greater than 3 in deep or (2) estimating flow with a ping pong ball (distance traveled / time = v). Instantaneous stream discharge (Q) was then calculated using corresponding stream channel measurements (i.e., depth and width). Stream temperature, dissolved oxygen (DO), pressure, pH, specific conductivity, and salinity were measured with a YSI Professional Plus Multiparameter Instrument. Standard pH solutions were used to calibrate YSI instruments prior to use. PME miniDO₂T Loggers II were placed in all streams excluding Painted Rocks (due to flow constraints) to continuously measure DO and temperature in 10-minute intervals for the entire study period.

Each week, water samples were collected for the analysis of the following stream constituents: total dissolved nitrogen (TDN), dissolved organic carbon (DOC), dissolved inorganic carbon (DIC), specific UV absorbance at 254 nm (SUVA₂₅₄), dissolved CO₂, δ¹³C-DOC, δ¹³C-DIC, δ¹³C-CO₂, and δ¹⁸O-H₂O. At the time of collection, all samples were filtered

through 0.45 μm syringe filters in the field. DIC samples were taken by injecting 1 mL of filtered stream water into prepared evacuated exetainers with 0.2 mL H_3PO_4 . To measure stream CO_2 concentrations, 30 mL of stream water was equilibrated with 30 mL of ambient air while submerged in the stream, and 15 mL of headspace from the syringe was added to a 12 mL exetainer. Field duplicates of all gas (including ambient air) and DIC samples were obtained. To analyze aquatic microbial communities, stream water was filtered until water could no longer exit through 0.22- μm Sterivex filters (MA, USA), biweekly—totaling five samples per site. Microbial sampling coincided with the biweekly field incubations described below.

All samples were stored on ice for transport back to Colorado College (Colorado Springs, CO). CO_2 , DIC, and DOC samples—which were processed within two weeks of collection—were promptly stored at 5°C, while isotope samples were frozen until analyses could be completed. Filters for DNA extraction and 16S ribosomal RNA (rRNA) gene sequencing were stored at -80°C.

Solute and DOC Analysis

The above solute samples and DOC quality analyses were completed during the summer of 2019 at Colorado College. TDN and DOC samples were analyzed on a Shimadzu TOC-L/TNM-L with standards using KHP and KNO_3 (0.1 to 10 mg L^{-1} C and N). Each sample's specific UV absorbance at 254 nm was measured using an Agilent UV-Vis Spectrometer. These values—when normalized to DOC concentrations (absorbance at 254 nm/[DOC]*100, in $\text{L mg C}^{-1} \text{ m}^{-1}$)—function as a proxy for DOC aromaticity (Weishaar *et al.*, 2003). Gas samples (ambient air and stream CO_2) were measured on an SRI-8610C gas chromatograph. Standard gases of 100 ppm, 1000 ppm, 10000 ppm CO_2 , and ambient lab air were run alongside samples for instrument calibration and equilibrium calculations.

Results from the above analyses were used to complete calculations for further study. Given the TDN and DIN (which consists of nitrite, nitrate, and ammonium) concentrations, dissolved organic nitrogen (DON) concentrations were determined ($\text{DON} = \text{TDN} - \text{DIN}$). The C:N of DOM was calculated using DOC and DON molar concentrations. Solute yields were calculated by dividing solute fluxes ($\text{flux} = [\text{solute}] * Q$) by respective watershed areas ($\text{yield} = \text{g m}^{-2} \text{ year}^{-1}$). DOC yields were also converted to grams of C year^{-1} and compared to estimates of grams of C exported from the soil using data from Jones (2020) to calculate DOC:soil C year^{-1} ratios.

Isotopic Analysis

Samples were sent to the Yale Analytical and Stable Isotope Center lab (Yale University, New Haven, CT, USA) for isotopic characterization. Analyses for $\delta^{13}\text{C}\text{-CO}_2$, $\delta^{13}\text{C}\text{-DOC}$, $\delta^{13}\text{C}\text{-DIC}$, and $\delta^{18}\text{O}\text{-H}_2\text{O}$ were performed on a Thermo DeltaPlus XP isotope ratio mass spectrometer. Carbon isotopic signatures indicated C sources to and within streams for CO_2 , DOC, and DIC. Additionally, $\delta^{18}\text{O}\text{-H}_2\text{O}$ measurements helped characterize differences in source waters and terrestrial processing prior to stream export.

Metabolism Measurements

In-situ three-hour dark incubations were completed according to Bogard *et al.* (2019) to measure planktonic respiration and indirectly assess DOC quality. These incubations were repeated five times throughout the summer at Sugar, Cabin, Pine, and Fourmile. Due to extremely low flows, incubations could not be conducted at Painted Rocks. At each site, stream water was added to a plastic five-gallon bucket until completely full to limit gas exchange with ambient air. Stream water in the bucket was clear of obvious aquatic plants, debris, litter, and sediment. A PME miniDO₂T Logger II was suspended in the bucket to record temperature and

DO concentrations every minute for 2.5-3 hours. While stream water incubated, the bucket remained submerged in flowing water as much as possible and covered with white plastic to ensure minimal temperature change from stream conditions.

The stream incubation mesocosm experiment measured DO uptake, the rate of which is representative of rapid mineralization of labile C in the water column (Bogard *et al.*, 2019). The difference in oxygen concentration over time ($\text{mg O}_2 \text{ L}^{-1} \text{ day}^{-1}$) in the bucket was calculated relative to DO saturation to account for changes in water temperature over the course of the incubation. However, since the bucket was not characteristic of stream conditions (i.e., it was often deeper and had no sediment), this rate demonstrates the relative bioavailability of C in the water column but not in-situ heterotrophic respiration rates (Bogard *et al.*, 2019). Molar ratios of $\text{O}_2:\text{DOC day}^{-1}$ were then quantified for each stream incubation by comparing the incubation respiration rate and respective stream water DOC concentrations given the volume of the bucket (18.93 L). This ratio serves as a proxy for stream DOC biodegradability.

Using the continuous 10-minute interval DO concentrations and temperature measurements from the miniDO₂T loggers, weekly whole stream metabolism metrics—net ecosystem production ($\text{NEP} = \text{GPP} - \text{ER}$), ecosystem respiration (ER), and gross primary productivity (GPP)—were calculated for Sugar, Cabin, Pine, and Fourmile following the inverse modeling method described by Hotchkiss & Hall (2014). Estimations of reaeration coefficients (k_{600}) were calculated based on stream temperature, the scaling equation, and the Schmidt number conversions for O₂ (Wanninkhof, 1992; Jähne *et al.*, 1987). Photosynthetically active solar radiation and barometric pressure characteristics used in this method were modeled based on latitude, longitude, and elevation data of the sites (Yard *et al.*, 1995; Colt, 2012). Metabolism metrics were estimated in R using the streamMetabolizer package (Appling *et al.*, 2018). In cases

where modeled GPP outputs were negative, values were corrected to zero given that negative productivity cannot occur.

Microbial Community Characterization

DNA extractions from the Sterivex filters were performed using Quick-DNA Fecal/Soil Microbe Kits (Zymo Research, CA, USA) in the fall of 2019 at the Wilkins Lab at Colorado State University (Fort Collins, CO). 16S rRNA gene sequencing and amplification was completed at Argonne National Laboratory (Lakewood, CO) on an Illumina HiSeq2500 sequencing system using the bacterial/archaeal primer set 515F/806R. This primer targets the V4 region and ensures maximum coverage of bacteria and archaea species and produces polymerase chain reaction (PCR) products of adequate length. Subsequent reads were grouped into exact sequence variants (ESVs) at 100% similarities via the QIIME 2 pipeline (QIIME2-2018.11). Sequences were archived in the NCBI sequence read archive under BioProject ID PRJNA659031.

Data Analysis

Statistical analysis and data visualization were completed using R Version 3.6.1. Pearson correlations were calculated to determine direction and strength of relationships between water quality measurements (including those of solutes, DOM quality, metabolism, and microbial community diversity metrics) and percent watershed burned, as well as other relevant variables such as water yield. This metric was also quantified for in-stream isotopes and qualitative stream variables. Isotopic measurements were considered temporally and compared to known signatures to determine sources. Two-sample t-tests were used to assess differences between streams draining burned or unburned catchments, and relationships with p-values less than or equal to

0.05 were considered statistically significant. Linear regression with a 95% confidence interval was used to assess relationships already identified as significant.

Alpha diversity, evenness, and richness metrics—Shannon’s Diversity Index (H), Simpson's Index (D), Pielou’s Evenness (J), and species richness—were calculated for further analysis of microbial communities within samples. Shannon’s Diversity Index combines both abundance and evenness measurements with 0 being the least diverse, as does Simpson’s Index (values = 0-1). Pielou’s evenness (values = 0-1) considers how close in numbers each species in the sample are with 1 being the most even. Species richness is based on how many different species are present in the sample.

The *vegan* (Oksanen *et al.*, 2007) and *phyloseq* (McMurdie & Holmes, 2013) packages in R were used to analyze differences in 16S rRNA gene datasets between streams and sampling dates. To assess differences in community composition between samples, Bray-Curtis dissimilarities were calculated. Bray-Curtis dissimilarity quantifies how dissimilar the microbial structures are. These values were plotted on a non-metric multidimensional scaling (NMDS) ordination to visualize differences in beta diversity between samples. The *envfit* function in R identified environmental variables and stream characteristics associated with microbial community composition shifts. The proximity of points to a vector illustrates a correlation, while vector length represents the strength of this relationship. In addition, analyses of similarity (ANOSIM) were computed to evaluate community differences between sites, burned and unburned watersheds, and burn extent.

RESULTS

Watershed Characterization

Stream discharge ($\text{ft}^3 \text{sec}^{-1}$) and water yield (mm year^{-1}) variability are the result of flow paths and catchment water export, which can both be altered by fire (Hallema *et al.*, 2017). At all five sites, peak discharge occurred by mid-June and declined throughout the study period, as temperature increased. This trend is expected as discharge of these montane streams is dominated by spring snowmelt. Additionally, discharge and water yield varied significantly among sites throughout the summer (Table 2; Fig. 2). The average discharge in the most extensively burned watershed (Fourmile) was the highest of the streams studied ($1.58 \pm 1.42 \text{ ft}^3 \text{sec}^{-1}$), and average discharge was relatively lower in streams draining unburned catchments (Sugar: $0.70 \pm 0.63 \text{ ft}^3 \text{sec}^{-1}$; Painted Rocks: $0.063 \pm 0.079 \text{ ft}^3 \text{sec}^{-1}$). Average water yields were significantly higher in burned ($41.99 \pm 34.33 \text{ mm year}^{-1}$) than unburned watersheds ($15.01 \pm 15.50 \text{ mm year}^{-1}$); however, this relationship ($p < 0.001$; Fig. 2) was largely driven by Fourmile's high (and variable) water yields ($56.56 \pm 51.09 \text{ mm year}^{-1}$).

Other stream parameters measured in the field did not consistently vary between sites, with fire history, or temporally. pH remained circumneutral in all streams and ranged from 7.11 to 7.90. Synoptic measurements of DO were similar among four of the stream sites, ranging from 71.5% to 81.4%, while DO measurements at Painted Rocks were significantly lower ($p = 0.01$) and ranged from 36.0% to 76.9%.

Stream Solutes

Dissolved Organic Matter

DOC concentrations in streams are linked to allochthonous and autochthonous processes and illustrate net differences in C sources and processing between watersheds. Although average

stream DOC concentrations did not vary with extent watershed burned, streams draining burned landscapes generally had lower DOC concentrations than those draining Ponderosa forest (Table 2). The average DOC concentration in Painted Rocks, an unburned watershed, was the highest among the sites studied ($5.66 \pm 0.89 \text{ mg C L}^{-1}$). The other unburned stream, Sugar, experienced the second lowest average DOC concentration ($1.63 \pm 0.38 \text{ mg C L}^{-1}$) among the five streams. Still, on average, DOC was lower at the three burned watersheds—Cabin, Pine, and Fourmile—than at Painted Rocks ($3.45 \pm 0.48 \text{ mg L}^{-1}$, $3.62 \pm 1.10 \text{ mg L}^{-1}$, and $1.62 \pm 0.43 \text{ mg L}^{-1}$ respectively). Lastly, DOC yields, DOC exports normalized to respective watershed areas, increased with burn extent (percent watershed burned) ($r = 0.31$, $p = 0.026$), largely due to greater water yields ($r = 0.76$, $p < 0.001$) in burned catchments (average values shown in Fig. 2).

Overall, SUVA_{254} values were significantly higher in unburned watersheds ($p < 0.001$) (Table 2) and decreased with percent burned ($r = -0.68$, $p < 0.001$; Fig. 3). SUVA_{254} values were as low as $2.30 \pm 0.38 \text{ L mg C}^{-1} \text{ m}^{-1}$ in the most extensively (82.36%) and severely burned (57.6%) site (Fourmile) and increased to $3.30 \pm 0.53 \text{ L mg C}^{-1} \text{ m}^{-1}$ in the least extensively (28%) and severely burned (14.9%) site (Cabin). In streams draining unburned landscapes, SUVA_{254} values over the summer were higher; Sugar averaged $3.37 \pm 0.75 \text{ L mg C}^{-1} \text{ m}^{-1}$, and Painted Rocks averaged $3.87 \pm 0.44 \text{ L mg C}^{-1} \text{ m}^{-1}$. Because SUVA_{254} values are a proxy for DOC aromaticity (Weishaar *et al.*, 2003), these results also imply that the C exported by burned watersheds was less aromatic (i.e., aromaticity and burn extent were negatively correlated). SUVA_{254} values, and therefore aromaticity of C, in streams draining burned watersheds did not fluctuate as much over the study period as compared to the DOM in streams draining unburned catchments. Within unburned watersheds, SUVA_{254} values increased over the course of the summer as discharge decreased ($r = -0.82$, $p < 0.001$) and experienced greater temporal

variability throughout the summer (Sugar: range = 2.37 L mg C⁻¹ m⁻¹; Painted Rocks: range = 1.49 L mg C⁻¹ m⁻¹) (Fig. 3). The temporal differences in SUVA₂₅₄ between burned and unburned watersheds suggest that stream DOC may be from different sources and/or processed differently due to shifts in flow paths.

DOC to DON ratios within streams are important to consider as they can be indicative of hydrologic flow paths and are often determined from soil C:N ratios (Aitkenhead-Peterson *et al.*, 2005). The ratio of DOC:DON was greatest in the unburned watershed Sugar (76.91 ± 53.34). Although the three streams in burned watersheds all had lower DOC:DON ratios than Sugar, DOC:DON was not significantly different between streams based on fire history.

DOC to soil C ratios illustrate how export of C from the watershed differs and is dependent on %C soil and water yield. This ratio was on average higher in burned watersheds (0.00063 ± 0.00048 year⁻¹) than in unburned watersheds (0.00027 ± 0.00043 year⁻¹) and increased with extent watershed burned ($r = 0.75$, $p = 0.05$; average values shown in Fig. 4).

$\delta^{13}\text{C-DOC}$

Conclusions can be made regarding C sources by comparing $\delta^{13}\text{C}$ signatures of DOC with known $\delta^{13}\text{C}$ values in the literature. The mean $\delta^{13}\text{C}$ in streams draining burned landscapes was $-30.28 \pm 0.95\text{‰}$ versus $-29.97 \pm 1.80\text{‰}$ in streams draining unburned landscapes (Table 3). Fourmile, an extensively burned watershed, experienced the most depleted $\delta^{13}\text{C-DOC}$ signal on average ($-30.93 \pm 0.97\text{‰}$), while Painted Rocks, an unburned watershed, experienced the most enriched $\delta^{13}\text{C-DOC}$ signal ($-29.06 \pm 1.08\text{‰}$). Stream $\delta^{13}\text{C-DOC}$ did not vary significantly with burn extent or over the course of the summer. At all five streams, $\delta^{13}\text{C-DOC}$ values remained steady throughout the study period (burned: CV = 0.03; unburned: CV = 0.06) and were not

correlated with discharge. The stable isotope signatures indicate consistent sources of organic matter from May to August.

Inorganic Carbon

Small headwater streams are typically supersaturated in CO₂; therefore, CO₂ is a significant contribution to the aquatic DIC pool (Campeau *et al.*, 2017; Finlay, 2003). CO₂ concentrations were significantly higher in streams draining unburned landscapes (1530.20 ± 819.72 ppm) than those draining burned landscapes (745.32 ± 236.28 ppm) (p = 0.0004). Average CO₂ concentrations ranged from 1945 ± 975 ppm in an unburned watershed (Painted Rocks) to 619 ± 87 ppm in the low extent burned watershed (Cabin) (Table 3). Moreover, CO₂ concentrations exhibited a negative relationship with discharge (r = -0.37, p = 0.01); stream concentrations increased over the course of the summer as water export decreased. CO₂ concentrations varied the most over the course of the summer in Painted Rocks, and unburned watersheds in general (Fig. 5). CO₂ concentrations were also positively correlated with stream DOC concentrations (r = 0.36, p = 0.01).

δ¹³C-DIC and δ¹³C-CO₂

The δ¹³C values of DIC and CO₂ reveal input sources to/within streams and subsequent C processing. δ¹³C-DIC signatures in streams incorporate δ¹³C-CO₂ from biogenic inputs and atmospheric evasion, as well as geogenic and photooxidative sources of inorganic C (Campeau *et al.*, 2017). Given that stream DIC and CO₂ are in equilibrium relative to pH and temperature, it was expected that δ¹³C-DIC and δ¹³C-CO₂ were significantly and positively correlated (r = 0.90, p < 0.001). On average, DIC and CO₂ were more enriched in ¹³C in burned than unburned sites (p < 0.001; Fig. 5). δ¹³C-DIC in unburned watersheds had an average value of -10.48 ± 1.16‰ as

compared to $-12.39 \pm 0.84\%$ in burned watersheds (Table 3). DIC was most enriched in ^{13}C at Cabin, a moderately burned watershed, ($-10.17 \pm 0.62\%$) and most depleted at Painted Rocks ($-12.94 \pm 0.80\%$). $\delta^{13}\text{C-CO}_2$ values reached $-16.61 \pm 1.76\%$ in burned watersheds, and like DIC, was most enriched in ^{13}C at Cabin ($-15.88 \pm 2.19\%$; Table 3). In unburned sites, $\delta^{13}\text{C-CO}_2$ averaged $-20.93 \pm 2.43\%$; Painted Rocks experienced the most depleted C signature as was the case with DIC ($-22.19 \pm 2.15\%$). Average summertime $\delta^{13}\text{C-DIC}$ and $\delta^{13}\text{C-CO}_2$ values additionally increased with percent burned ($r = 0.54$, $p < 0.001$; Fig. 5).

In burned and unburned streams, $\delta^{13}\text{C}$ signatures of DIC and CO_2 became more depleted throughout the summer, and $\delta^{13}\text{C-DIC}$ was correlated with both discharge ($r = 0.56$, $p < 0.001$) and temperature ($r = -0.24$, $p = 0.10$). There was greater temporal variability in unburned watersheds (Fig. 5), and relationships with discharge were stronger in these streams as well (Sugar: $r = 0.87$, $p = 0.001$; Painted Rocks: $r = 0.81$, $p = 0.005$). In all watersheds except for Cabin, $\delta^{13}\text{C-DIC}$ increased with discharge, yet this relationship was only significant at Fourmile, a severely burned site ($r = 0.71$, $p = 0.02$). Similarly, $\delta^{13}\text{C-CO}_2$ was only significantly associated with discharge in unburned streams ($r = 0.74$, $p = 0.004$) and Fourmile ($r = 0.73$, $p = 0.03$).

Nitrogen

As observed in other studies (Rhoades *et al.*, 2011), burn extent and severity largely predicted variation in summer stream nitrogen (N) concentrations and the species of dissolved N present. Fourmile had the highest average TDN concentration ($0.58 \pm 0.36 \text{ mg N L}^{-1}$) over the summer, while Sugar had the lowest average ($0.059 \pm 0.064 \text{ mg N L}^{-1}$) (Table 2). However, TDN concentrations in Fourmile were greatly determined by DIN (average = $0.43 \pm 0.19 \text{ mg L}^{-1}$), which was dominated by nitrate (average = $0.42 \pm 0.19 \text{ mg N L}^{-1}$), while DON dominated the dissolved nitrogen pools in the other four streams (all DIN averages $< 0.1 \text{ mg L}^{-1}$).

$\delta^{18}\text{O}\text{-H}_2\text{O}$

$\delta^{18}\text{O}\text{-H}_2\text{O}$ measurements can be indicative of flow paths, as well as overall hydrology, and are dependent on temperature (Gat, 2010). On average, $\delta^{18}\text{O}\text{-H}_2\text{O}$ was the most depleted in Sugar ($-14.34 \pm 0.23\text{‰}$), an unburned site. H_2O was most enriched in ^{18}O in unburned site Painted Rocks ($-12.52 \pm 0.11\text{‰}$) and the most severely burned watershed Fourmile ($-13.22 \pm 0.29\text{‰}$) (Fig. 6). Excluding the smallest watershed, Painted Rocks, $\delta^{18}\text{O}\text{-H}_2\text{O}$ values increased with extent watershed burned ($r = 0.58$, $p < 0.001$). Significant relationships were not found between $\delta^{18}\text{O}\text{-H}_2\text{O}$ and temperature or discharge, which were also both strongly correlated with burn extent. However, $\delta^{18}\text{O}\text{-H}_2\text{O}$ tended to increase through the study period from May to August ($r = 0.42$).

Stream Metabolism

In Situ Incubations

Incubation rates—which describe rates of labile C mineralization in streams—did not vary significantly between burned and unburned watersheds or between individual sites. Average rates also varied between streams. Both the maximum ($5.66 \text{ mg O}_2 \text{ L}^{-1} \text{ day}^{-1}$) and minimum ($0.61 \text{ mg O}_2 \text{ L}^{-1} \text{ day}^{-1}$) rates occurred at Cabin, indicating large temporal variability within a single site. Although similar amounts of variability occurred at the other four sites, incubation rates were not statistically correlated with temperature or discharge. Furthermore, incubation rates were not statistically correlated with any solute measured during the experiment (e.g., TDN, DOC, SUVA_{254} , $\delta^{13}\text{C}\text{-DIC}$).

Dissolved C Bioavailability

From bucket incubation oxygen consumption rates, molar ratios of $\text{O}_2\text{:DOC day}^{-1}$ were calculated as a measure of stream DOC bioavailability. The ratio varied temporally at all sites.

Unlike incubation rates, which did not vary with disturbance history, O₂:DOC day⁻¹ ratios were greater in unburned watersheds on average (0.65 ± 0.28) as compared to burned watersheds (0.45 ± 0.31) (Fig. 7). However, Fourmile, the most extensively burned watershed, had the highest C bioavailability on average among the five sites (0.77 ± 0.29), while Cabin, another burned watershed, experienced the lowest average ratio (0.29 ± 0.22). Among streams draining burned watersheds, O₂:DOC day⁻¹ ratios significantly increased with extent watershed burned ($r = 0.66$, $p = 0.007$; Fig. 7). These ratios were negatively correlated with $\delta^{13}\text{C}$ -DOC ($r = -0.50$, $p = 0.03$; Fig. 8), and this relationship was most significant at Cabin ($r = 0.99$, $p = 0.0014$) and Sugar ($r = 0.90$, $p = 0.04$).

Modeled Daily Metabolism

Weekly whole stream metabolism estimations from in-situ O₂ sensor measurements illustrated that all five streams were dominated by ecosystem respiration (ER) and had low levels of gross primary productivity (GPP). Net ecosystem productivity was negative at all sites throughout the summer, as expected. ER was not significantly different between sites, with average ER estimates ranging from 1.49 ± 0.36 g O₂ m⁻² day⁻¹ to 1.89 ± 0.42 g O₂ m⁻² day⁻¹. ER was positively correlated with water yield across all sites ($r = 0.50$, $p = 0.001$).

Rates of GPP were often higher in burned watersheds than unburned. However, significantly higher rates of GPP occurred at unburned site Sugar, which averaged 1.22 ± 0.40 g O₂ m⁻² day⁻¹, while moderately burned site Cabin averaged 0.055 ± 0.17 g O₂ m⁻² day⁻¹. Negative outputs occurred in 37% of the estimates, most often in Cabin and Pine, suggesting the model (Hotchkiss & Hall, 2014) did not capture all variability in the data. Mineralization of DOC rates calculated from the bucket incubations were not correlated with the modeled rates of whole stream respiration (ER).

Microbial Communities

Diversity Metrics

Community diversity in aquatic microbial communities may be influenced by factors such as OM pools and temperature (Zeglin, 2015). Accordingly, species richness, Shannon's Diversity Index, Simpson's Index, and Pielou's evenness varied according to burn severity, although not always significantly. Aquatic microbial communities from burned and unburned watersheds had similar Pielou's evenness. Still, burned watersheds experienced slightly higher Pielou's evenness on average (range = 0-1) (0.85 ± 0.087) than aquatic microbial communities from unburned watersheds (0.81 ± 0.078). Species richness was highest in Pine (2417 ± 432 ESVs), a moderately burned watershed, and lowest in Fourmile (1517 ± 493 ESVs), a severely burned watershed. Microbial species richness decreased with extent burned, but this was not a statistically significant relationship ($r = -0.20$, $p = 0.35$). No significant differences were found for Shannon's Diversity and Simpson's Index values between burned and unburned aquatic microbial communities ($p > 0.05$).

Both microbial community composition and diversity often show temporal heterogeneity as well as connections to ecosystem-scale responses (Zeglin, 2015). Exact sequence variants (ESVs) were most similar within sites (ANOSIM, Sugar vs. Painted Rocks vs. Cabin vs. Pine vs. Fourmile, 0.784 , $p < 0.001$) and were also statistically similar between sites with similar fire history and burn extent (ANOSIM, burned vs. unburned watersheds, 0.558 , $p < 0.001$; ANOSIM, % burned, 0.546 , $p < 0.001$). When plotted on an NMDS ordination, Bray-Curtis dissimilarity values had a relatively low stress value (0.113), indicating a fair fit of the ordination (Fig. 9). Bray-Curtis dissimilarities illustrated significant differences between microbial assemblage composition based on site location and extent burned (Fig. 9). These results further reinforce

relationships between site characteristics and fire history. Microbial communities differentiated based on SUVA₂₅₄ values and TDN concentrations across the burn gradient. Greater SUVA₂₅₄ were correlated with the microbial communities at unburned sites (Fig. 3), and greater TDN concentrations (Table 2) were correlated with the microbial communities at burned sites. Bray-Curtis dissimilarities confirmed differences between microbial communities as explained by temporal variables including hydrological characteristics (e.g., average velocity), inorganic C isotope signatures, and diversity metrics (e.g., species richness).

Differences in Relative Abundance

Differences in the relative abundances of microbial phyla between burned and unburned sites can demonstrate how the stream microbiome is altered by fire and fire severity. The six most abundant phyla in streams draining burned and unburned watersheds (in order of decreasing relative abundance in burned watersheds) included Proteobacteria, Bacteroidetes, Cyanobacteria, Verrucomicrobia, Patescibacteria, and Actinobacteria. On average, streams draining burned landscapes contained greater relative abundances of Proteobacteria, Bacteroidetes, Actinobacteria, and Verrucomicrobia than unburned streams, which contained greater relative abundances of Cyanobacteria and Patescibacteria (Table 4; Fig. 10a). Relative abundances of Bacteroidetes, Verrucomicrobia, and Actinobacteria were significantly different between burned and unburned sites ($p = 0.008, 0.001, 0.002$, respectively; Table 4). Also, certain families of Actinobacteria—Intrasporangiaceae, Nocardiaceae, and Mycobacteriaceae—were more abundant in streams from burned catchments (Fig. 10b).

DISCUSSION

Changes to C Export and Processing

Understanding linkages between terrestrial and aquatic environments following wildfire is important for assessing its legacy on carbon partitioning, and thus carbon sequestration, across the landscape. Seventeen years after the Hayman Fire, changes to the terrestrial environment persist. Vegetation significantly differed compared to unburned sites, resulting in shifting hydrology and a characteristically different soil and soil microbiome (Bonwell, 2020; Jones, 2020). Fire alteration of vegetation and soil properties plays a significant role in OM quality (González-Pérez *et al.*, 2004; Knicker, 2007; Fornwalt & Kaufmann, 2014) and amount of C exported to streams (Moody & Martin, 2001; Rhoades *et al.*, 2018). In burned watersheds, SOM appears to be more bioavailable (Jones, 2020) and was more likely to be laterally exported to streams than in unburned catchments (Bonwell, 2020); therefore, C likely has a shorter residence time in these terrestrial landscapes (Fig. 11).

Export of DOC is largely related to terrestrial C cycling and balance, and burned systems are no exception (Aitkenhead-Peterson *et al.*, 2005). Amendments to the antecedent environmental conditions, which may be driven by fire, impact aquatic nutrient export in response to changes in temperature (Xu & Saiers, 2010), discharge, seasonality (Dawson *et al.*, 2008), and overall DOM mobilization from the watershed (Raymond & Saiers, 2010). Wildfire often leads to elevated water export (Fig. 2) due to decreased vegetative cover and decreased evapotranspiration (Hallema *et al.*, 2017), which occurred in the three burned watersheds here. Given that water yield was highest in burned catchments (Cabin, Pine, and Painted Rocks), DOC export was also greater in the moderately burned watersheds (Cabin and Pine) than the unburned watersheds (Sugar and Painted Rocks), consistent with previous studies (Rhoades *et al.*, 2018).

Moreover, DOC export was greater per unit C in the watershed (Fig. 4), suggesting that a combination of factors, including increased water yield and reduced %C in burned soils (Jones, 2020), may lead to greater transport of C from terrestrial to aquatic environments.

Differences in the quality of C exported can also be understood in terms of riparian and upland vegetation differences between sites and the slow recovery of vegetation after fire (Rhoades *et al.*, 2018). Lower SUVA₂₅₄ values indicate that DOM was less aromatic (Weishaar *et al.*, 2003) in streams draining burned landscapes as compared to intact forests. In addition, pyrogenic carbon (highly aromatic C produced during fire) in soils did not vary statistically between burned and unburned watersheds 15 years following the Hayman Fire (Barnes & Gilbertson, 2018, unpublished data), potentially due to fire history and the residence time of black carbon. However, previous studies examining stream DOC exports, typically occurring within a decade following fire, report higher stream SUVA₂₅₄ values with the influx of pyrogenic and other condensed aromatic C compounds (Santos *et al.*, 2016; Olefeldt *et al.*, 2013; Chow *et al.*, 2019). Although SUVA₂₅₄ values remained elevated 14 years post-Hayman Fire in moderately burned watersheds (Chow *et al.*, 2019), our results indicate that DOC export moved away from the aromatic C pools that typically dominate streams following fire (Fig. 3). Shifts in vegetation and thus C inputs (e.g., lignin, cellulose, etc.) after fire likely drive this pattern. Minimal forest recovery in 17 years led to the dominance of herbaceous cover (i.e., grasses and forbs, $19.0 \pm 10.1\%$ and $10.9 \pm 5.39\%$ cover respectively, Jones, 2020) in burned regions of the Hayman burn scar. In contrast, woody plants (i.e., Ponderosa pine and shrubs) dominate the unburned landscapes, contributing aromatic inputs of lignin, as compared to the less aromatic cellulose inputs within burned catchments.

The interaction between hydrology and vegetation also drives carbon dynamics in these watersheds and its lateral transport to streams. Isotopic analysis (Table 3) reveals that both inorganic and organic C pools in all streams were predominantly terrestrially derived, as expected for small headwater streams (Vannote *et al.*, 1980; Palmer *et al.*, 2001; Hotchkiss *et al.*, 2018). For example, $\delta^{13}\text{C}$ -DIC signatures ($-11.24 \pm 1.35\text{‰}$) generally fell between values for atmospheric CO_2 ($\sim 8\text{‰}$, Finlay & Kendall, 2007) and CO_2 from soil respiration ($-14.11 \pm 4.19\text{‰}$, Jones, 2020). Integration of the atmospheric CO_2 signature occurred as small streams are typically supersaturated in CO_2 , and this disequilibrium allows for the rapid evasion of CO_2 from stream surfaces to the atmosphere (Öquist *et al.*, 2009). $\delta^{13}\text{C}$ -DIC values additionally reflect that stream inorganic C pools are influenced by terrestrial SOM decomposition (i.e., CO_2 from microbial respiration; Fig. 12), similar to other studies, which only emphasizes the importance of quantifying and qualifying DIC fluxes (Finlay, 2003; Campeau *et al.*, 2017). Additionally, DIC and CO_2 were more enriched in ^{13}C in burned sites than in unburned sites, reflecting the more isotopically enriched, albeit smaller, soil CO_2 pools in burned watersheds (Jones, 2020). Lastly, average $\delta^{13}\text{C}$ -DOC values ($-30.28 \pm 0.95\text{‰}$) in burned streams were more depleted than that of catchment SOM ($\sim 24\text{‰}$, Jones, 2020) (Fig. 5), implying preferential retention of ^{13}C following leaching (Hagedorn *et al.*, 2004) or different sources of DOC in the aquatic systems, such as bacterial communities.

Fire history likely influences in-situ processes, which respond to fluctuations in C stocks (Langenheder *et al.*, 2010) and changing abiotic conditions. At all sites, as dissolved CO_2 concentrations increased throughout the summer, $\delta^{13}\text{C}$ - CO_2 signatures became more depleted (Fig. 5), suggesting increased rates of respiration as temperatures rose (Aerts, 1997; Fierer *et al.*, 2005). During the beginning of the study period in May and June, $\delta^{13}\text{C}$ - CO_2 closely reflected the

C signature of the CO₂ respired from soil at both burned and unburned catchments. However, the $\delta^{13}\text{C}$ -CO₂ signature became noticeably more depleted over the course of the summer at unburned sites, trending toward the signatures of the watershed SOM and stream DOC (Fig. 5). This trajectory indicates a shift in flow path (or contributing source area) and/or differences in in-situ sources and processing between sites over time. For example, greater rates of evapotranspiration in forests could lead to greater temporal variability in flow paths, which was also evident in trends in $\delta^{18}\text{O}$ -H₂O (Fig. 6).

Oxidation of stream DOM may impact aquatic C stocks in streams draining unburned watersheds to a greater extent given that it was more aromatic and less processed than the DOM in streams draining burned landscapes (Fig. 3). According to measurements from in situ bucket incubations, DOM in burned streams was also less bioavailable (Fig. 7), and bacterial production may be reduced in these sites due to lower species richness in streams draining burned landscapes. These data may also explain the lack of temporal signals within burned streams as compared to unburned streams where DOM was more bioavailable. Furthermore, OM bioavailability and $\delta^{13}\text{C}$ -DOC were negatively correlated (Fig. 8), illustrating a potential relationship between C availability to microbial communities and C character (Wiegner *et al.*, 2005; Kaplan *et al.*, 2008). Although SOM in burned soils was more bioavailable (Jones, 2020), C may be less bioavailable in burned streams as this C was processed more within the terrestrial landscape or because the SOM pool in burned soils was significantly smaller than that of unburned soils, and thus more likely to be processed before reaching the stream.

Shifts in the Stream Microbiome

Stream microbiomes were distinct from each other and were distinguishable by fire history. It is surprising that the aquatic microbiomes appear to be more distinct depending on site

and burn extent as well as more responsive to environmental factors like C quality—as illustrated by how close samples cluster to each other and plotted vectors (Fig. 9)—than the soil microbiomes of these watersheds (Jones, 2020). Soil microbiomes did not distinguish between sites and only differentiated based on fire history (see Fig. 9 in Jones, 2020).

Many factors may explain these shifts in community structure, including biogeochemical (e.g., DOM supply, Judd *et al.*, 2006) or environmental characteristics (e.g., pH, Fierer *et al.*, 2007). However, the NMDS ordination used to quantify dissimilarities between microbial community samples illustrates that the microbial composition changed with shifts in hydrology (e.g., velocity), organic matter quality (SUVA₂₅₄), and inorganic carbon and nitrogen pools ($\delta^{13}\text{C-DIC}$, TDN) (Fig. 9). The fact that SUVA₂₅₄ values and $\delta^{13}\text{C-DIC}$ were correlated with shifts in microbial assemblage implies that C supply exerts some control on the stream microbiome. Given the importance of DOC as the largest pool of organic matter in fluvial systems (Kaplan *et al.*, 2008) and as a source of C to stream microorganisms, fire indirectly drives differences in microbiome composition by altering DOC export and C quality.

The C pool and character select for different microbial taxa over a relatively short time period (Judd *et al.*, 2006). For example, microbiomes in streams draining unburned watersheds exhibited the most inter-site variability, which also experienced greater variability in water yield, DOC export, SUVA₂₅₄, and ^{13}C isotopic signatures than streams draining burned watersheds. Streams draining burned watersheds exported greater DOC concentrations per unit C in hillslope soils (Fig. 4), which could likewise yield differences in microbial communities (Eiler *et al.*, 2003). In essence, these shifts in DOM quality and export post-fire may elucidate the observed differences in stream microbiomes both in the short-term and long-term (Bernhardt & Likens,

2002; Crump *et al.*, 2003; Eiler *et al.*, 2003; Judd *et al.*, 2006; Strickland *et al.*, 2009; Zeglin, 2015).

Although aquatic C stocks may influence the microbiome, aquatic microbial communities and other in-stream processes also affect dissolved organic and inorganic C stocks. Different relative abundances of certain microbial phyla possibly inform the changes in C quality via increased microbial transformation of C, preferential degradation of less aromatic C compounds, or a greater effect of photodegradation on aromatic DOM within burned watersheds (Olefeldt *et al.*, 2013; Hotchkiss *et al.*, 2018). Heterotrophic taxa within the Bacteroidetes, Actinobacteria, and Verrucomicrobia phyla were statistically more abundant in burned watersheds (Table 4; Fig. 10) and may all play key roles in the degradation of organic matter (Thomas *et al.*, 2011; Cardman *et al.*, 2014; Chen *et al.*, 2016). While it is challenging to predict functional properties of different taxa identified through marker gene analyses, it is likely that there are overlaps in the ecological functions of microorganisms within these phyla that result in minor differences in the aquatic C stock.

Actinobacteria—including three specific families (Fig. 10b)—that were more abundant in both burned soils (Jones, 2020) and streams draining those landscapes (Fig. 10) are associated with the degradation of aromatics, including pyrogenic carbon in burned watersheds (Khodadad *et al.*, 2011; Chen *et al.*, 2016). Soil pyrogenic C amendments have been shown to drive an increase in the relative abundance of bacteria in the phylum of Actinobacteria following fire (Khodadad *et al.*, 2011). Given that greater amounts of pyrogenic C no longer remain in these watersheds this many years post-fire (Barnes & Gilbertson, 2018, unpublished data) but elevated Actinobacteria relative abundances persist (Fig. 9), the legacy of fire on burned landscapes appears to extend to the microbial communities beyond that of certain C modifications. Although

the conditions which enriched this phylum have diminished, the persisting higher abundances of Actinobacteria in streams draining burned landscapes potentially illustrates the ability of wildfire to disturb ecosystems to a new steady state.

Even though microbial communities differed based on fire history, differences in in-stream processes (e.g., ER) did not reflect these community shifts. Given that ecosystem respiration did not vary significantly between streams, but the microbial community structure did, functional redundancy—the ecological phenomenon when different taxa assume the same functional niche—may exist in these systems. Therefore, fire seems to govern stream systems in ways that are either not detectable in measurements of ecological functioning, or that the communities function differently but still result in same process rates when considered at the community level (Allison & Martiny, 2008).

CONCLUSION

Overall, the legacy of wildfire on these landscapes extends to carbon sources, subsequent processing, and microbial community composition in both terrestrial and aquatic environments 17 years after the Hayman Fire (Fig. 11). C partitioning and residence times have been altered in the terrestrial environment due to differences in the dominant vegetation (Jones, 2020). Moreover, shifts in hydrology and the soil C stock post-fire led to increased lateral export of C to aquatic systems in burned landscapes. However, although allochthonous sources dominated these small headwater streams, C was less bioavailable and qualitatively different once transported. Compared to streams draining unburned landscapes, in-situ processing may have a smaller effect on burned systems, and microbial assemblages experienced significant shifts nearly two decades after the disturbance. Although differences in C character inevitably impact aquatic microbial composition, differences in stream microbiomes may alter aquatic C stocks and transformation as well. Thus, wildfire potentially forces a positive feedback loop between C export and aquatic microbiomes by furthering C alteration (Fig. 11). The altered recovery dynamics for both vegetation and microbiomes suggest that the Hayman Fire has shifted these systems to a new steady state in the face of severe disturbance (Stevens-Rumann *et al.*, 2018).

Understanding how C is partitioned across watersheds like those studied here is pertinent when considering the global C balance in response to climate change. Expanding knowledge on the lateral export of C to inland waters is often disregarded in these assessments, and microbial mediation of C is typically ignored in small streams. As such, it is valuable to address these issues following disturbances like fire, which are expected to increase in both frequency and severity in the coming decades (Westerling *et al.*, 2006; Abatzoglou & Williams, 2016).

TABLES AND FIGURES

	Sugar	Painted Rocks	Cabin	Pine	Fourmile
Latitude	39.30476	39.08249	39.23229	39.24051	39.24257
Longitude	-105.1818	-105.1039	-105.302	-105.2819	-105.2311
Area (km²)	36.2	4.5	24.0	11.1	24.9
% Burned	0	0	28	53.7	82.3
% Severely Burned	0	0	14.9	34.1	57.6

Table 1. Burn site characteristics, including latitude and longitude data for site locations. The watershed area of each site is listed as well as the percent of the watershed that burned and was severely burned.

	Sugar	Painted Rocks	Cabin	Pine	Fourmile
% Burned	0	0	28	53.7	82.3
Water Yield (mm year⁻¹)	17.35 ± 15.70	12.68 ± 15.77	24.19 ± 14.68	45.21 ± 19.34	56.56 ± 51.09
DOC (mg C L⁻¹)	1.63 ± 0.38	5.66 ± 0.89	3.448 ± 0.49	3.62 ± 1.10	1.62 ± 0.43
DOC Yield (g C m⁻² year⁻¹)	30.22 ± 31.54	74.36 ± 99.71	83.92 ± 51.93	173.46 ± 109.47	95.84 ± 89.60
SUVA₂₅₄ (mg C⁻¹ m⁻¹)	3.37 ± 0.75	3.87 ± 0.44	3.30 ± 0.53	2.97 ± 0.32	2.30 ± 0.38
TDN (mg N L⁻¹)	0.06 ± 0.06	0.48 ± 0.15	0.29 ± 0.09	0.18 ± 0.09	0.69 ± 0.35
TDN Yield (g N m⁻² year⁻¹)	1.51 ± 3.22	4.89 ± 5.97	6.00 ± 3.99	8.05 ± 7.54	26.59 ± 15.14

Table 2. Solute concentrations and quality measurements. Averages and standard deviations for water yields (mm year⁻¹), DOC and TDN concentrations (mg L⁻¹) and yields (g m⁻² year⁻¹), and SUVA₂₅₄ values for each watershed are provided.

	Sugar	Painted Rocks	Cabin	Pine	Fourmile
% Burned	0	0	28	53.7	82.3
$\delta^{13}\text{C-DOC}$ (‰)	-30.88 ± 1.95	-29.06 ± 1.08	-30.11 ± 0.82	-29.80 ± 0.73	-30.93 ± 0.97
$\delta^{13}\text{C-DIC}$ (‰)	-11.83 ± 1.23	-12.94 ± 0.80	-10.17 ± 0.62	-10.72 ± 0.79	-10.54 ± 1.05
$\delta^{13}\text{C-CO}_2$ (‰)	-19.66 ± 2.08	-22.19 ± 2.15	-15.88 ± 2.19	-16.42 ± 1.32	-17.46 ± 1.52
CO₂ (ppm)	1116 ± 294	1945 ± 795	619 ± 87	719 ± 138	885 ± 331
$\delta^{13}\text{C-SOM}$ (‰)	-24.30 ± 0.95	-24.33 ± 0.42	-24.20 ± 0.36	-24.27 ± 1.01	-24.47 ± 0.49
$\delta^{13}\text{C-CO}_2$, Soil Respiration (‰)	-14.85 ± 1.72	17.34 ± 2.01	-14.6 ± 0.95	-14.07 ± 0.89	-13.68 ± 0.67

Table 3. Isotopic analysis of aquatic and terrestrial C sources. Average ^{13}C signatures and standard deviations for DOC, DIC, and CO_2 , compared to CO_2 concentrations (ppm) and ^{13}C signatures from the watershed (SOM and CO_2 , Jones, 2020) for each catchment are provided.

Phylum	Average Relative Abundance (burned)	Average Relative Abundance (unburned)	p-value
Proteobacteria	0.394 ± 0.080	0.385 ± 0.098	0.806
Bacteroidetes	0.195 ± 0.061	0.129 ± 0.049	0.008*
Cyanobacteria	0.0841 ± 0.092	0.180 ± 0.147	0.104
Verrucomicrobia	0.0783 ± 0.027	0.0400 ± 0.012	0.0001*
Patescibacteria	0.0420 ± 0.029	0.0514 ± 0.036	0.5166
Actinobacteria	0.0517 ± 0.037	0.0148 ± 0.004	0.002*

Table 4. Average microbial relative abundances ± standard deviation for the six most abundant phyla found in all sites studied by burn history. Differences were tested with two-sample t-tests, and significance was defined as $p \leq 0.05$ (as indicated by *).

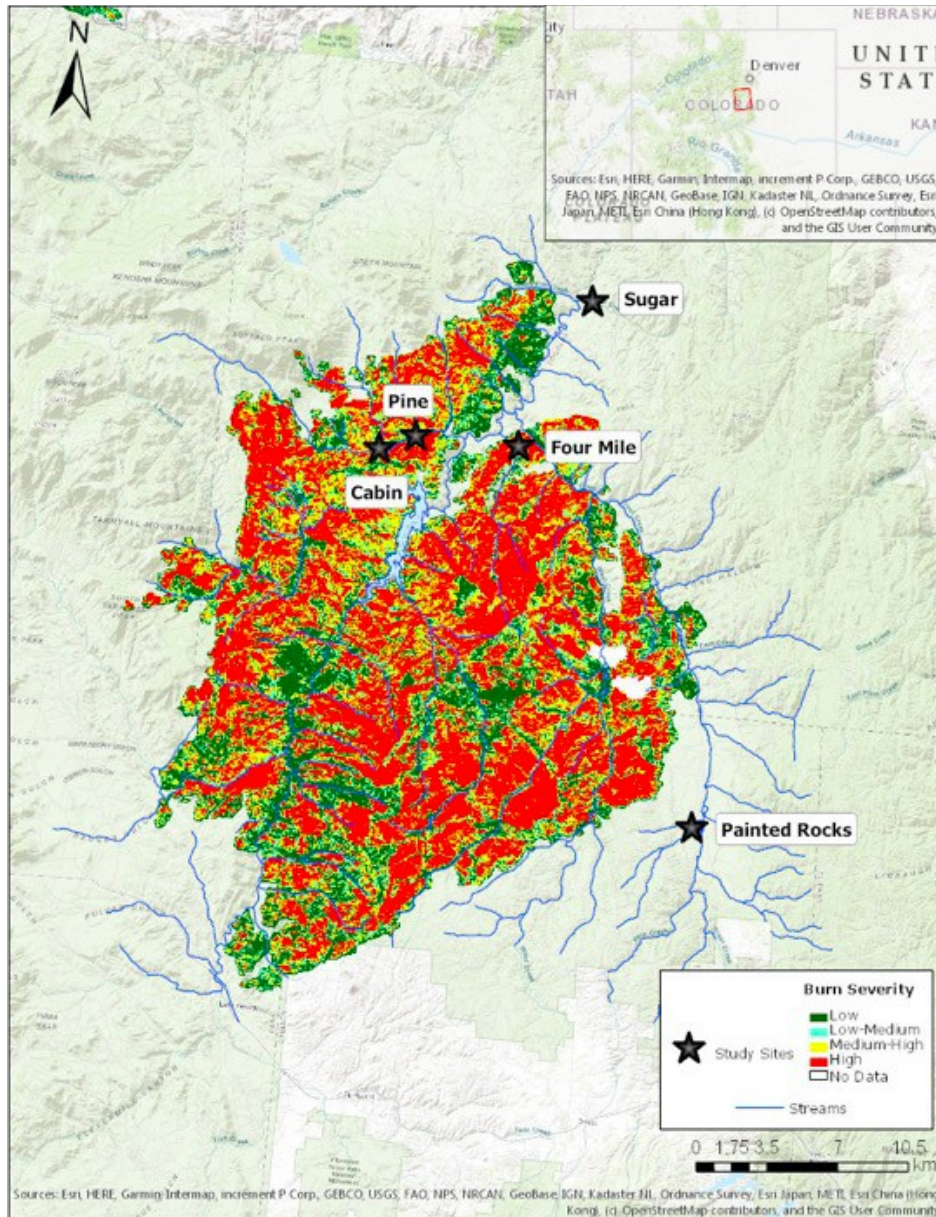


Figure 1. Map of study site locations (indicated with black star and name label) and burn extent of 2002 Hayman and Schoonover fires. Based on the Monitoring Trends in Burn Severity (MTBS) scale, burn severity is shown low (green) to high (red). Map created in ArcGIS Pro. *Figure courtesy of Cheristy Jones.*

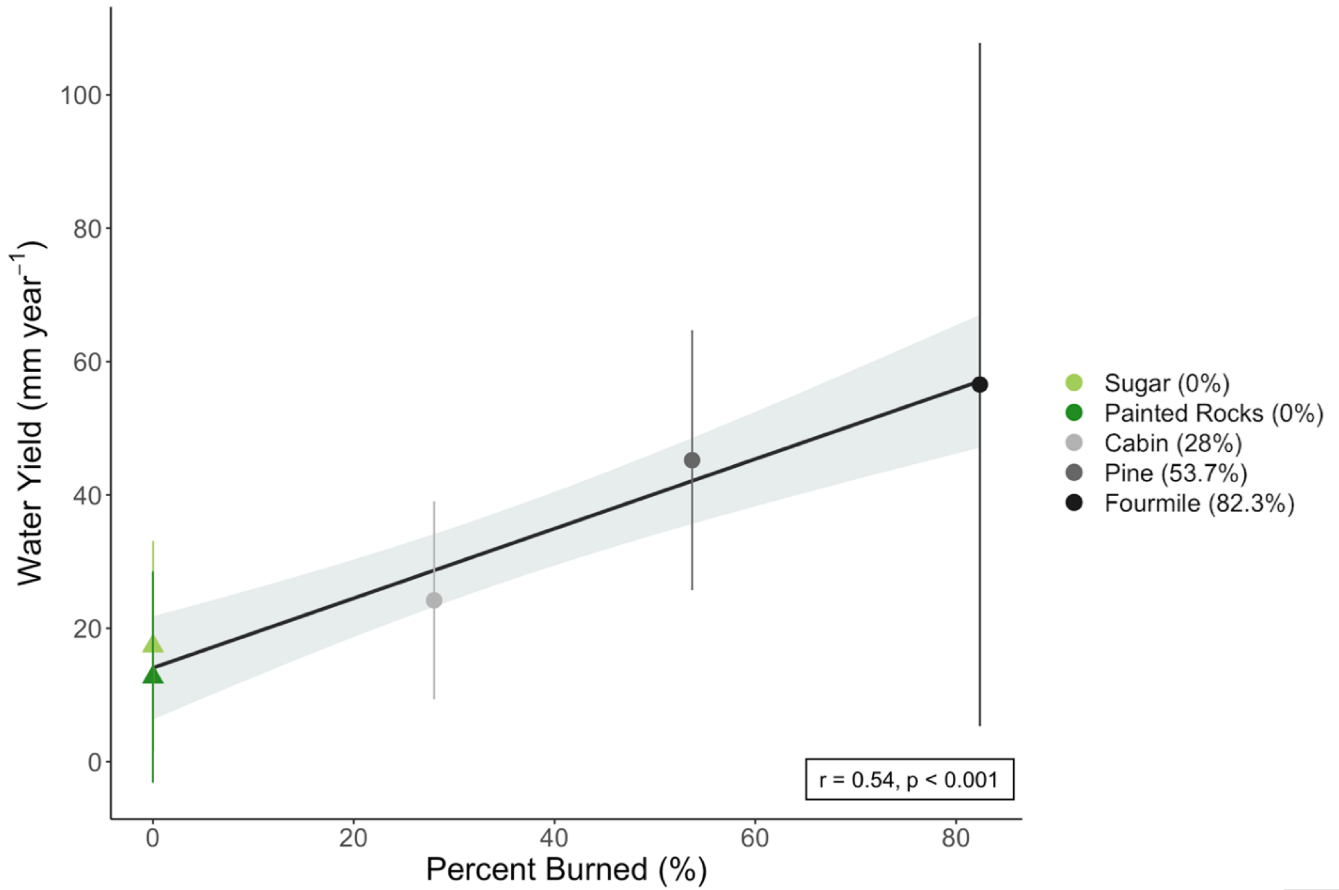


Figure 2. Average water yields (mm year⁻¹) by site based on percent watershed burned (%) with line of best fit. Gray coloration indicates the 95% confidence interval, and error bars are representative of \pm standard deviation. Circle points correspond to burned watersheds, and triangle points correspond to unburned watersheds. Water yield increased significantly with percent burned.

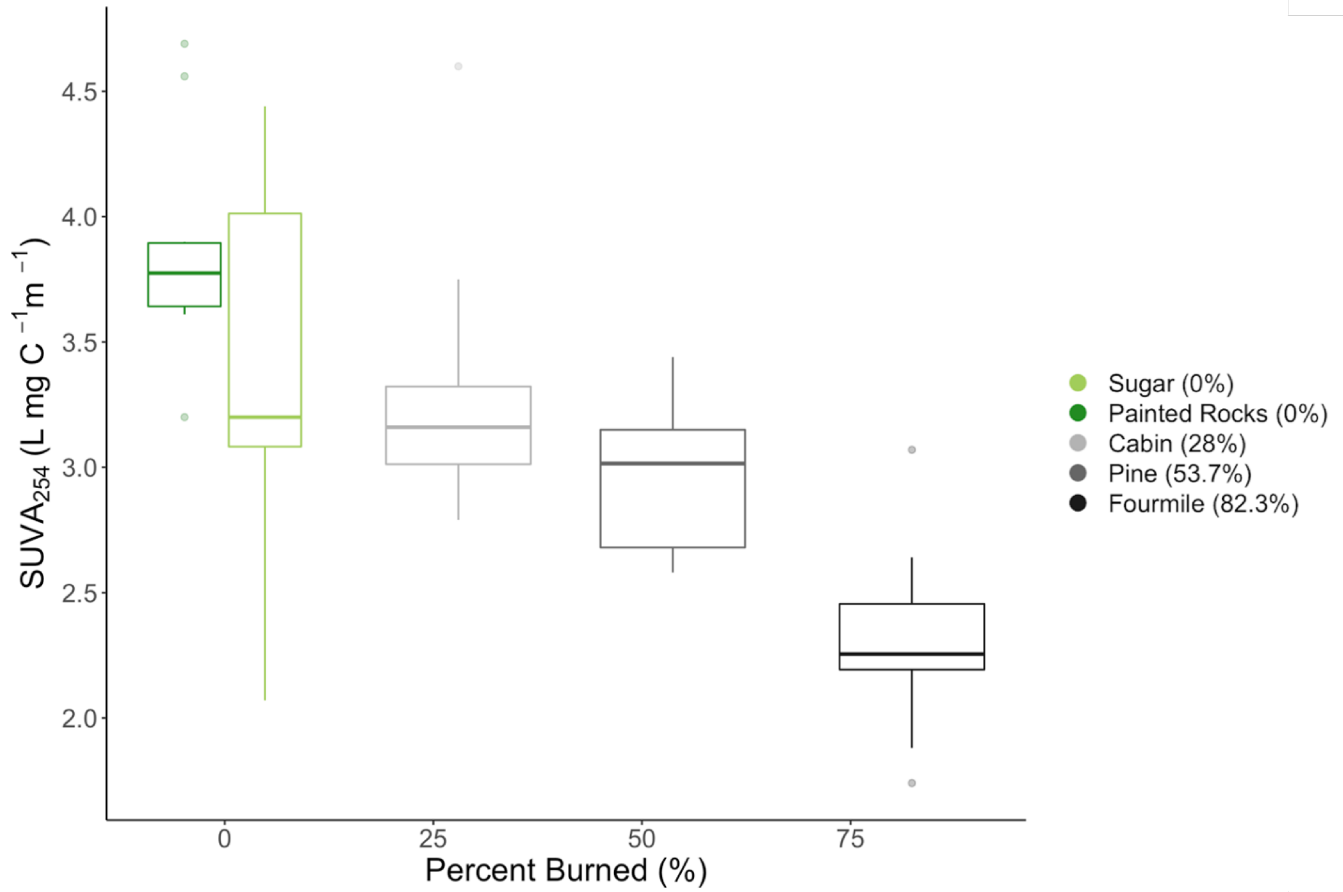


Figure 3. Box plot of SUVA₂₅₄ (L mg C⁻¹ m⁻¹) values according to percent watershed burned (%). SUVA₂₅₄ significantly decreased with percent burned ($r = -0.68$, $p < 0.001$), indicating that aromaticity also was highest in unburned watersheds and lowest in severely burned watersheds (Weishaar *et al.*, 2003).

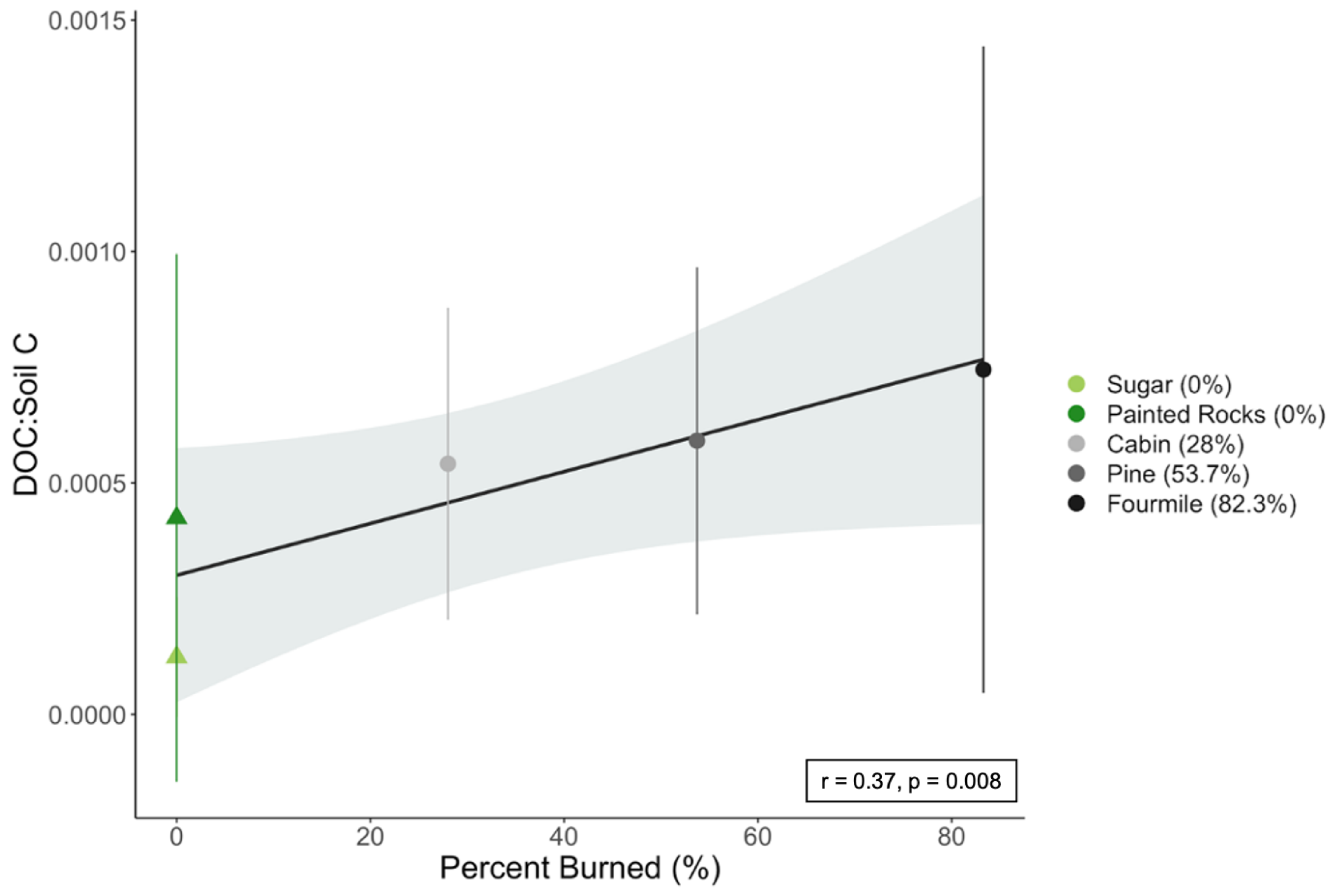


Figure 4. DOC:soil C (year^{-1}) ratios by percent watershed burned (%) with line of best fit. Gray coloration indicates the 95% confidence interval, and error bars are representative of \pm standard deviation. Circle points correspond to burned watersheds, and triangle points correspond to unburned watersheds. DOC to soil C ratios shed light on how watershed C export differs and is dependent on %C soil and water yield. Average ratios increased with extent watershed burned ($r = 0.37$, $p = 0.008$).

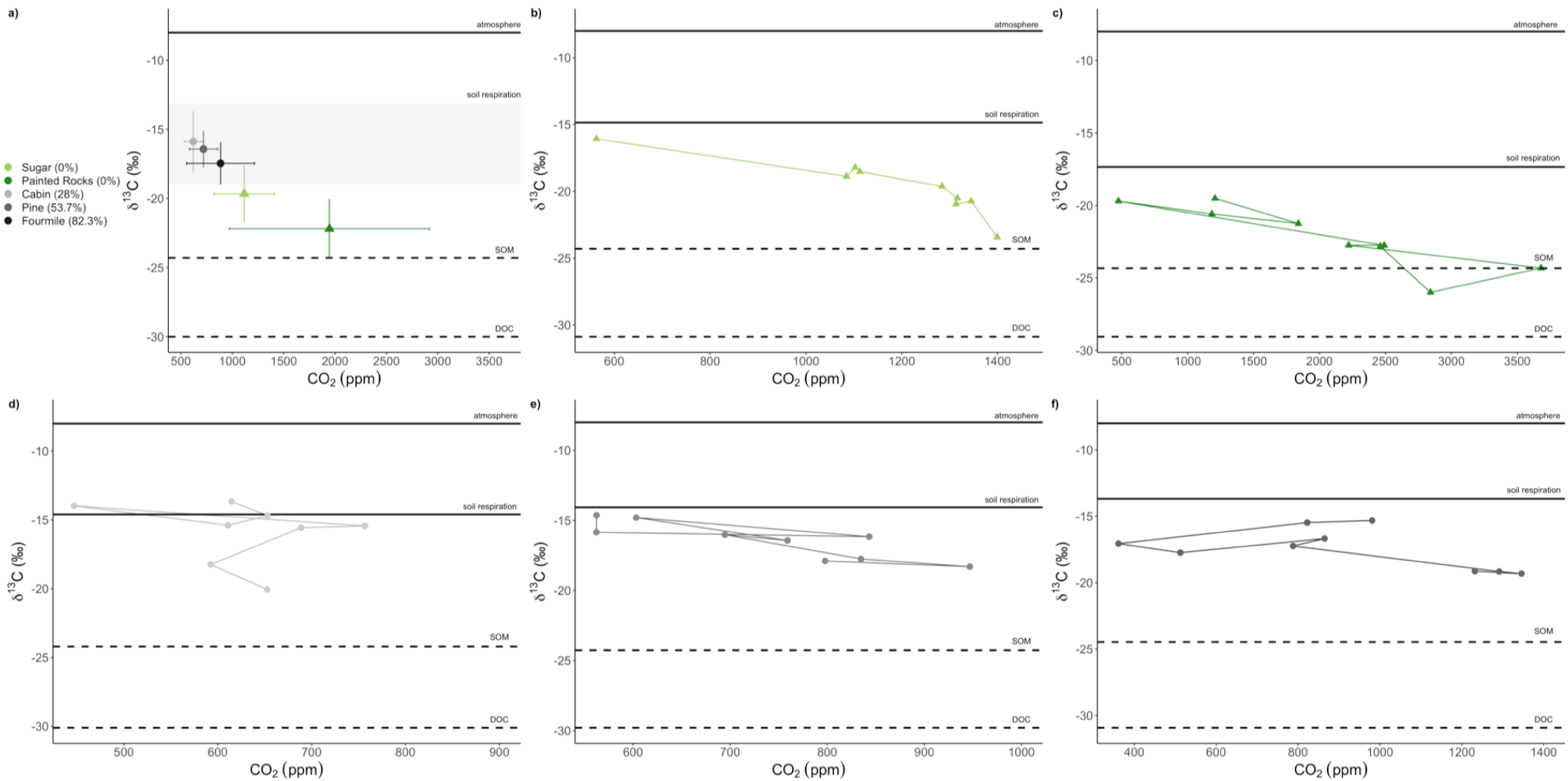


Figure 5. $\delta^{13}\text{C-CO}_2$ (‰) values compared to CO_2 concentrations (ppm). Circle points correspond to burned watersheds, and triangle points correspond to unburned watersheds. The solid black line represents average $\delta^{13}\text{C-CO}_2$ (atm) ($\sim -8\text{‰}$, Finlay & Kendall, 2007). Additionally, the two dashed lines represent average $\delta^{13}\text{C-SOM}$ and $\delta^{13}\text{C-DOC}$ values respectively, which may serve as sources to processes that produce CO_2 . These lines or regions are indicated as these C sources may be potential terrestrial sources of CO_2 . (a) The shaded grey region represents the range of values of $\delta^{13}\text{C-CO}_2$ (soil respiration) in all five watersheds. Plotted points are indicative of an average $\delta^{13}\text{C-CO}_2$ and average CO_2 concentrations in each site. Error bars are representative of \pm standard deviation and illustrate the temporal variation in these variables, which was greatest in the unburned watersheds for CO_2 concentration and $\delta^{13}\text{C-CO}_2$. (b-f) The solid black line at a more depleted C signature represents the average $\delta^{13}\text{C-CO}_2$ (soil respiration) value in a single watershed. Plotted points are connected temporally to demonstrate how CO_2 concentrations generally increase as $\delta^{13}\text{C-CO}_2$ becomes more depleted over the course of the summer. Unburned sites possibly reflect different in-situ sources or processing of C as the summer progresses.

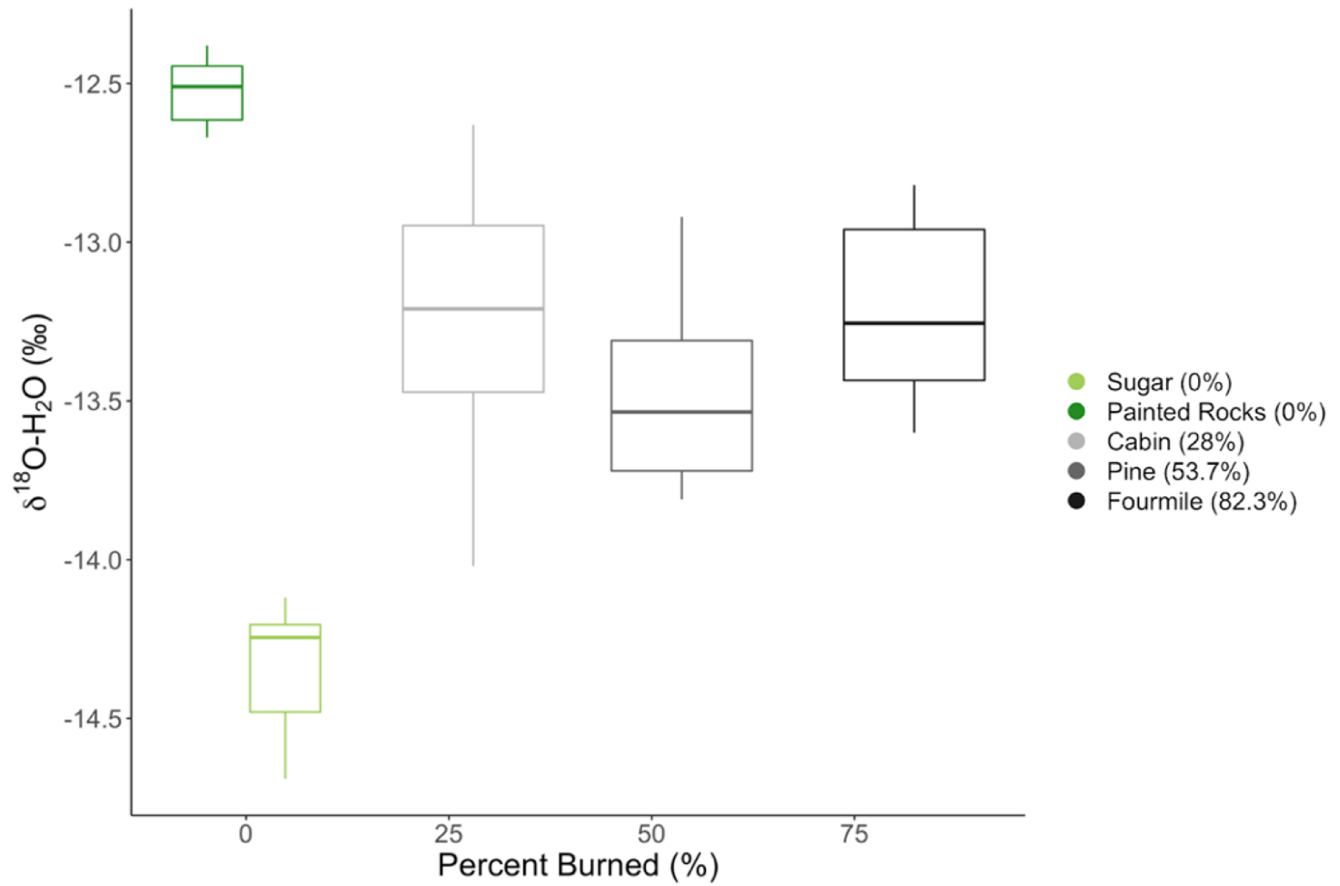


Figure 6. Box plot of $\delta^{18}\text{O-H}_2\text{O}$ (‰) according to percent watershed burned (%). $\delta^{18}\text{O-H}_2\text{O}$ values can be indicative of overall hydrology (Gat, 2010). When excluding the smallest watershed, Painted Rocks (an unburned site), $\delta^{18}\text{O-H}_2\text{O}$ values increased with extent watershed burned ($r = 0.58$, $p = 0.001$), thus becoming less depleted.

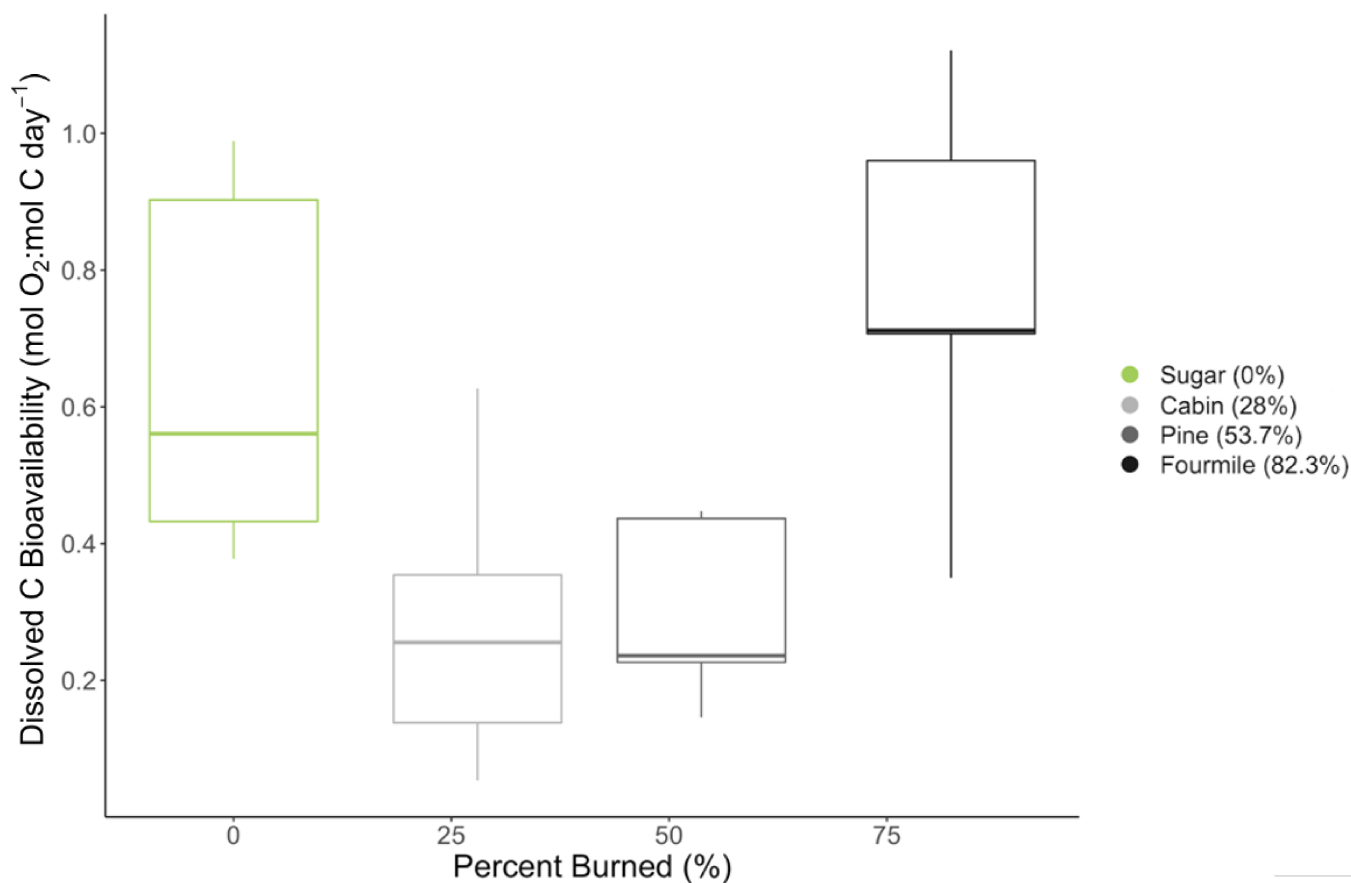


Figure 7. Box plot of dissolved C bioavailability ($\text{mol O}_2:\text{mol DOC day}^{-1}$) according to percent watershed burned (%). Measurements were not taken at the unburned site, Painted Rocks, due to extremely low flows. In general, $\text{O}_2:\text{DOC day}^{-1}$ ratios were greater in unburned watersheds on average as compared to burned watersheds, but all four sites experienced significant temporal variation. However, this temporal variation was more prominent in the unburned stream, Sugar. Also, among the three streams draining burned landscapes, $\text{O}_2:\text{DOC day}^{-1}$ ratios significantly increased with extent watershed burned ($r = 0.66$, $p = 0.007$).

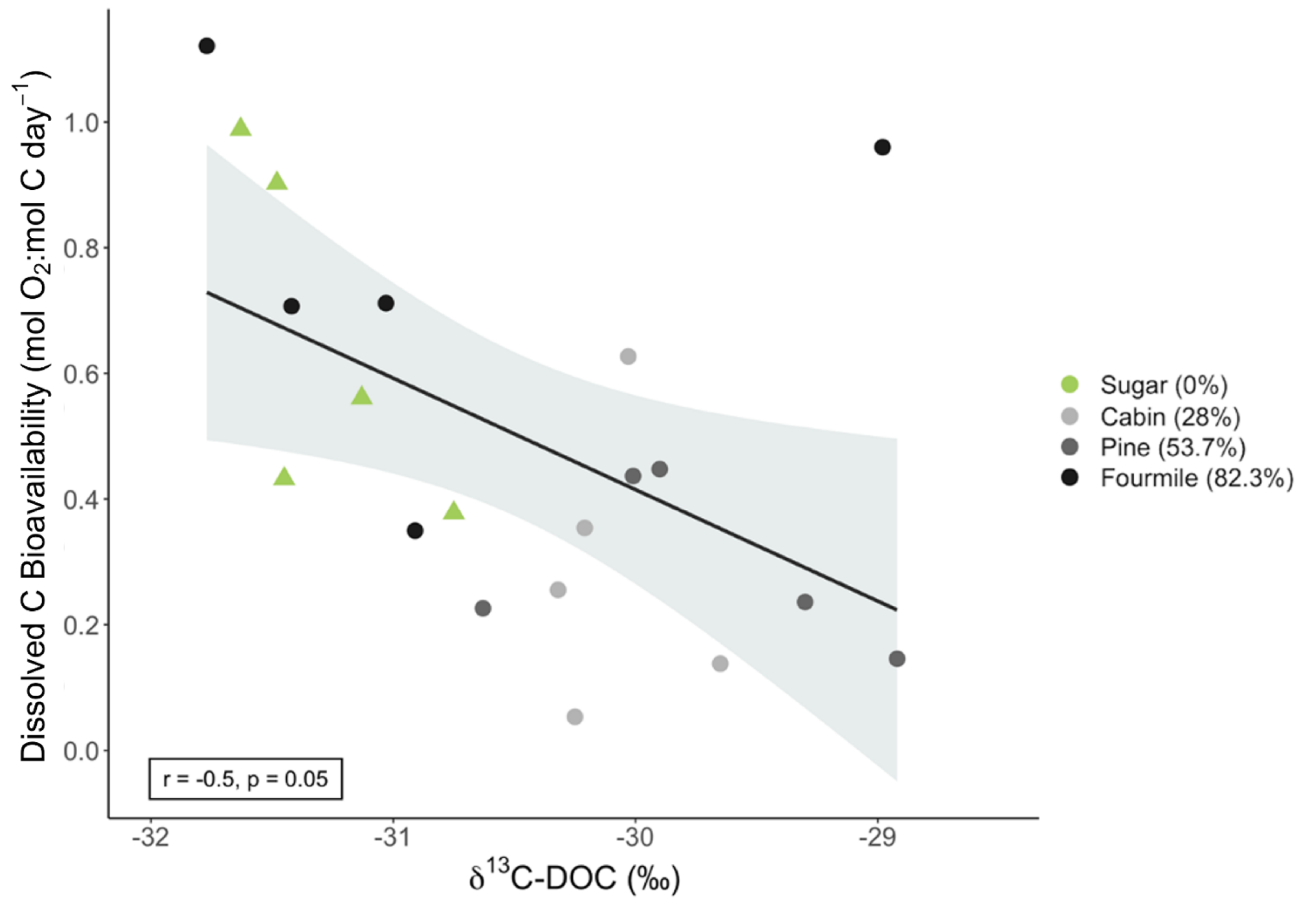


Figure 8. Dissolved C bioavailability (mol O₂:mol DOC day⁻¹) compared to δ¹³C-DOC with line of best fit. Measurements were not taken at the unburned site, Painted Rocks, due to extremely low flows. Gray coloration indicates the 95% confidence interval, and error bars are representative of ± standard deviation. Circle points correspond to burned watersheds, and triangle points correspond to unburned watersheds. Dissolved C bioavailability was negatively correlated with δ¹³C-DOC ($r = -0.50$, $p = 0.03$). This relationship was most significant at Cabin ($r = 0.99$, $p = 0.0014$) and Sugar ($r = 0.90$, $p = 0.040$).

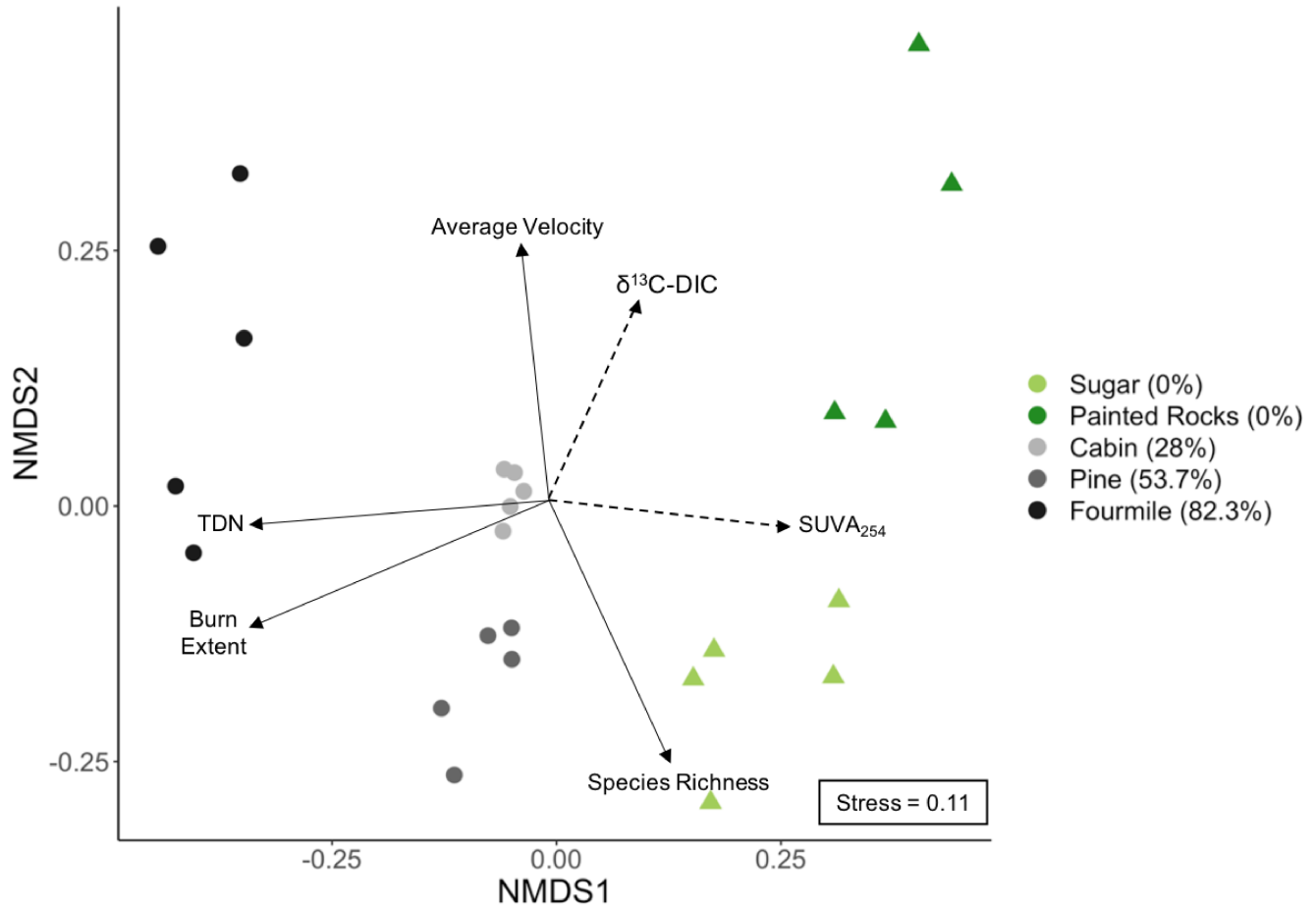


Figure 9. Non-metric multidimensional scaling (NMDS) ordination of exact sequence variants (ESVs) to assess beta diversity in aquatic microbial communities by watershed. Circle points correspond to burned watersheds, and triangle points correspond to unburned watersheds. The low stress value indicates a good fit of the ordination. Samples that are more similar (based on Bray-Curtis dissimilarities) are plotted closer together. Samples were most similar to those from the same stream (ANOSIM: 0.784, $p < 0.001$). Samples were also more similar to those with similar fire history or burn extent (ANOSIM, burned vs. unburned watersheds, 0.558, $p < 0.001$; ANOSIM, % burned, 0.546, $p < 0.001$). Vectors illustrate variables that were correlated with differences between samples. Solid vectors have a p -value < 0.05 , whereas dashed vectors have a p -value > 0.05 . The proximity of points to a vector illustrates a correlation, while vector length represents the strength of this relationship. As such, stream chemistry and burn extent appear to be associated with the microbial communities in the five streams studied here.

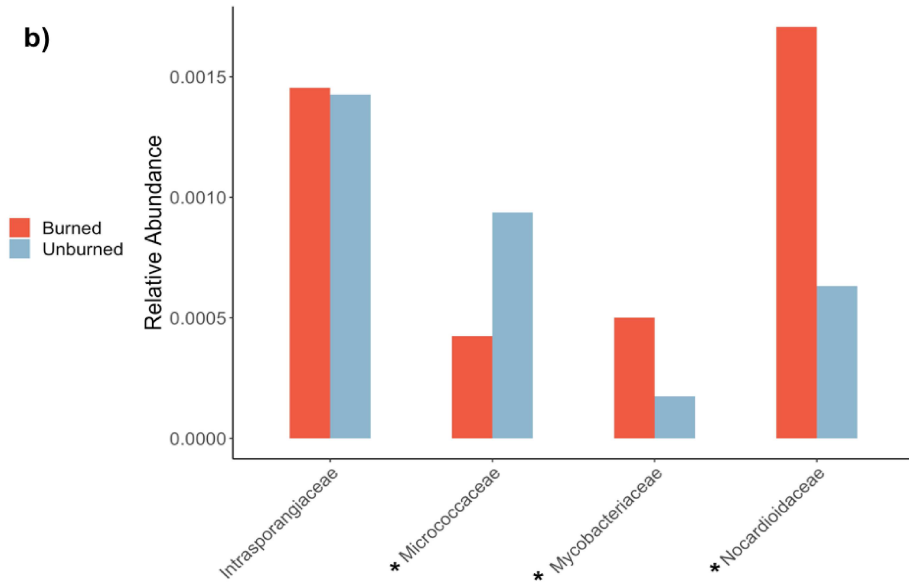
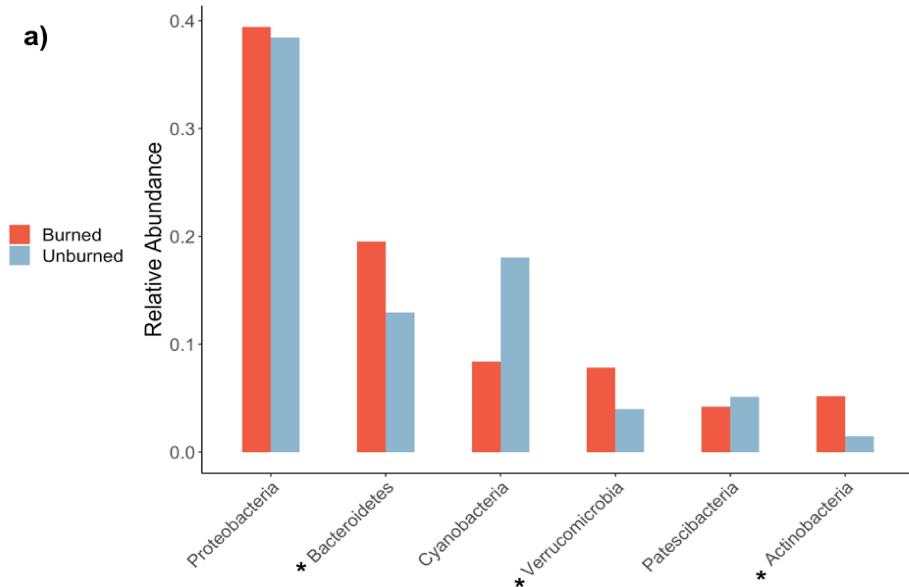


Figure 10. Relative abundances for ecologically relevant microbial taxa present in streams draining both burned and unburned landscapes. Stars next to phyla or family indicate that the difference in relative abundance between burned and unburned watersheds was statistically significant ($p \leq 0.05$). (a) Average relative abundances between burned and unburned watersheds for the six most abundant phyla across streams. Proteobacteria, Bacteroidetes, Verrucomicrobia, and Actinobacteria were more abundant in burned watersheds, whereas Cyanobacteria, and Patescibacteria were more abundant in unburned watersheds. (b) Average relative abundances between burned and unburned watersheds for four families of the Actinobacteria phyla that may be related to the degradation of aromatics. Three out of four of these families were more abundant in burned watersheds, excluding Micrococcaceae.

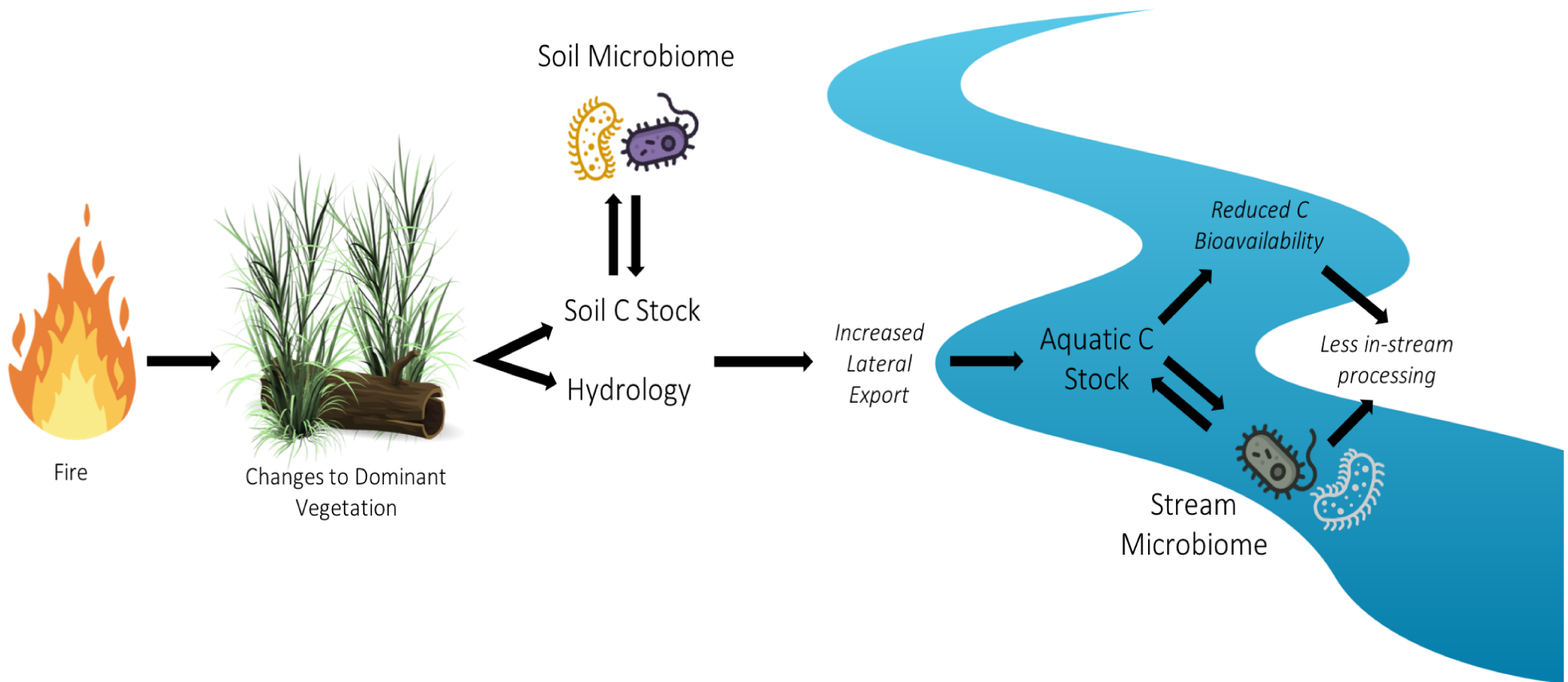


Figure 11. The effects of wildfire on C partitioning and processing across a montane landscape. Wildfire drastically affects the dominant vegetation present in a watershed, and the burned watersheds studied here were predominantly covered by grasses and forbs. Such changes to vegetation directly affect the soil and soil C stock; at these sites, soil C was more bioavailable (Jones, 2020), and C experienced shorter residence times (Bonwell, 2020). Soil microbial communities both influence this C pool and have also shifted in structure in response to SOM quality. Together, these impacts to the watershed led to increased lateral export of terrestrial C to aquatic systems. DOC export was elevated as a result and less bioavailable compared to that of streams draining unburned landscapes. C was qualitatively different in these watersheds (as evidenced in $SUVA_{254}$ and $\delta^{13}C$ values), potentially leading to diminished in-stream processing. Additionally, changes to the aquatic C stock may shift the stream microbial assemblage, and microbial processes can further determine DOC character.

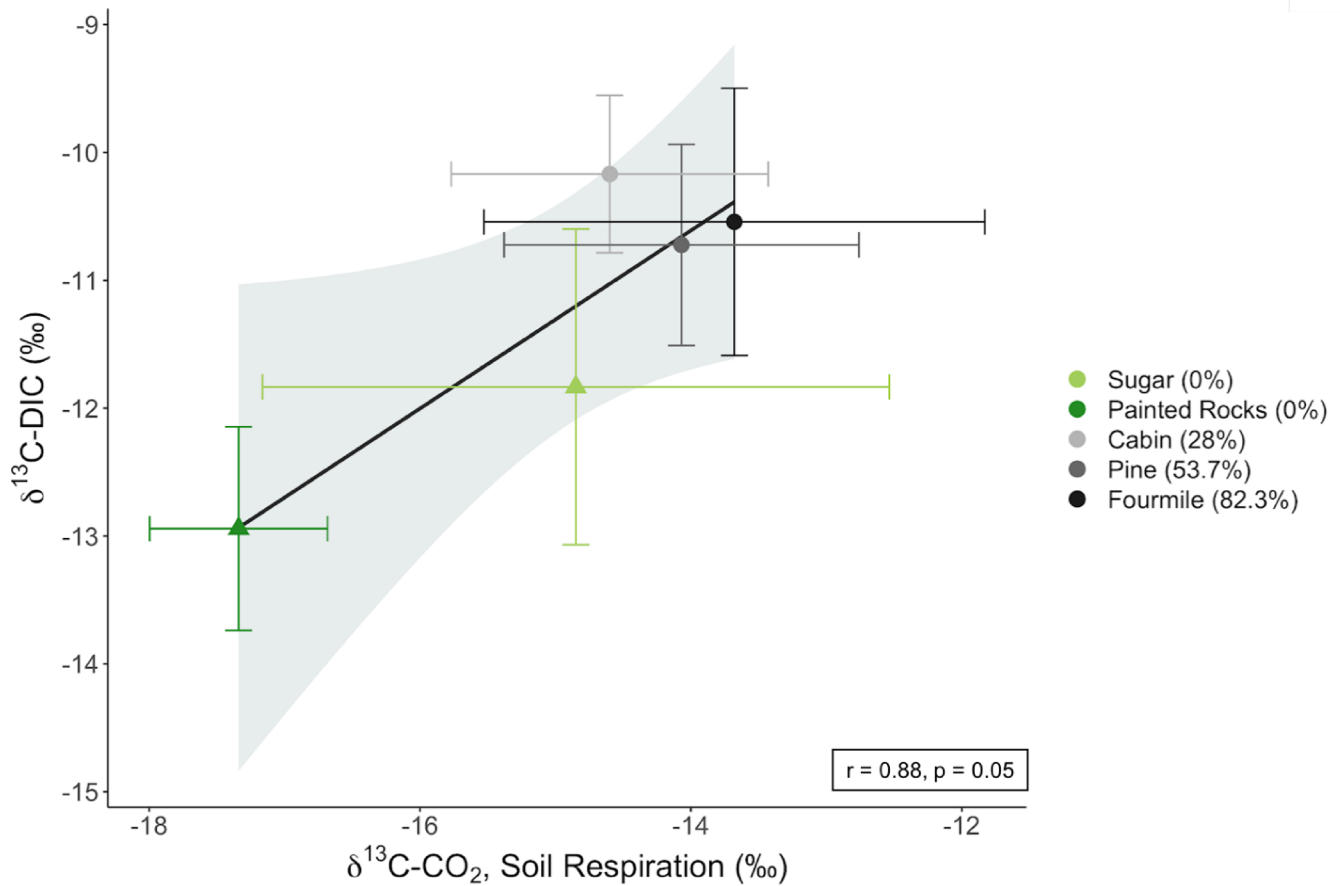


Figure 12. Average stream $\delta^{13}\text{C-DIC}$ (‰) values compared to average $\delta^{13}\text{C-CO}_2$ (‰) respired through soil microbial respiration in each watershed with line of best fit. Gray coloration indicates the 95% confidence interval, and error bars are representative of \pm standard deviation. Circle points correspond to burned watersheds, and triangle points correspond to unburned watersheds. Stream $\delta^{13}\text{C-DIC}$ values were correlated with $\delta^{13}\text{C-CO}_2$ from soil respiration ($r = 0.88, p = 0.05$). This relationship illustrates the influence of terrestrial sources on the aquatic DIC pool.

REFERENCES

- Abatzoglou, J. T., & Williams, A. P. (2016). Impact of anthropogenic climate change on wildfire across western US forests. *Proceedings of the National Academy of Sciences*, *113*(42), 11770-11775.
- Abella, S. R., & Fornwalt, P. J. (2015). Ten years of vegetation assembly after a North American mega fire. *Global change biology*, *21*(2), 789-802.
- Aerts, R. (1997). Climate, leaf litter chemistry and leaf litter decomposition in terrestrial ecosystems: a triangular relationship. *Oikos*, 439-449.
- Aitkenhead-Peterson, J. A., Alexander, J. E., & Clair, T. A. (2005). Dissolved organic carbon and dissolved organic nitrogen export from forested watersheds in Nova Scotia: Identifying controlling factors. *Global Biogeochemical Cycles*, *19*(4).
- Allison, S. D., & Martiny, J. B. (2008). Resistance, resilience, and redundancy in microbial communities. *Proceedings of the National Academy of Sciences*, *105* (Supplement 1), 11512-11519.
- Appling, A. P., Hall Jr., R. O., Yackulic, C. B., & Arroita, M. (2018). Overcoming equifinality: Leveraging long time series for stream metabolism estimation. *Journal of Geophysical Research: Biogeosciences*, *123*(2), 624-645.
- Bernhardt, E. S., & Likens, G. E. (2002). Dissolved organic carbon enrichment alters nitrogen dynamics in a forest stream. *Ecology*, *83*(6), 1689-1700.
- Betts, E. F., & Jones Jr, J. B. (2009). Impact of wildfire on stream nutrient chemistry and ecosystem metabolism in boreal forest catchments of interior Alaska. *Arctic, Antarctic, and Alpine Research*, *41*(4), 407-417.
- Bogard, M. J., Johnston, S. E., Dornblaser, M. M., Spencer, R. G., Striegl, R. G., & Butman, D. E. (2019). Extreme rates and diel variability of planktonic respiration in a shallow sub-arctic lake. *Aquatic Sciences*, *81*(4), 60.
- Bonwell, C. (2020). Impact of severe fire on stream carbon fluxes and metabolism across a mesic montane landscape. Undergraduate Thesis. Colorado College.
- Bowman, D. M., Balch, J. K., Artaxo, P., Bond, W. J., Carlson, J. M., Cochrane, M. A., ... & Pyne, S. J. (2009). Fire in the Earth system. *Science*, *324*(5926), 481-484.
- Bryant, B., McGrew, L. W., & Wobus, R. A. (1981). *Geologic map of the Denver 1 x 2 quadrangle, north-central Colorado*. US Geological Survey.
- Buma, B., Brown, C. D., Donato, D. C., Fontaine, J. B., & Johnstone, J. F. (2013). The impacts of changing disturbance regimes on serotinous plant populations and communities. *BioScience*, *63*(11), 866-876.
- Buma, B., Poore, R. E., & Wessman, C. A. (2014). Disturbances, their interactions, and cumulative effects on carbon and charcoal stocks in a forested ecosystem. *Ecosystems*, *17*(6), 947-959.
- Butman, D., Stackpole, S., Stets, E., McDonald, C. P., Clow, D. W., & Striegl, R. G. (2016). Aquatic carbon cycling in the conterminous United States and implications for terrestrial carbon accounting. *Proceedings of the National Academy of Sciences*, *113*(1), 58-63.

- Campeau, A., Wallin, M. B., Giesler, R., Löfgren, S., Mörth, C. M., Schiff, S., ... & Bishop, K. (2017). Multiple sources and sinks of dissolved inorganic carbon across Swedish streams, refocusing the lens of stable C isotopes. *Scientific Reports*, 7(1), 1-14.
- Cardman, Z., Arnosti, C., Durbin, A., Ziervogel, K., Cox, C., Steen, A. D., & Teske, A. (2014). Verrucomicrobia are candidates for polysaccharide-degrading bacterioplankton in an arctic fjord of Svalbard. *Applied and environmental microbiology*, 80(12), 3749-3756.
- Certini, G. (2005). Effects of fire on properties of forest soils: a review. *Oecologia*, 143(1), 1-10.
- Chen, P., Zhang, L., Guo, X., Dai, X., Liu, L., Xi, L., ... & Huang, Y. (2016). Diversity, biogeography, and biodegradation potential of actinobacteria in the deep-sea sediments along the southwest Indian ridge. *Frontiers in microbiology*, 7, 1340.
- Chow, A. T., Tsai, K. P., Feghel, T. S., Pierson, D. N., & Rhoades, C. C. (2019). Lasting Effects of Wildfire on Disinfection By-Product Formation in Forest Catchments. *Journal of Environmental Quality*, 48(6), 1826-1834.
- Cole, J. J., Prairie, Y. T., Caraco, N. F., McDowell, W. H., Tranvik, L. J., Striegl, R. G., ... & Melack, J. (2007). Plumbing the global carbon cycle: integrating inland waters into the terrestrial carbon budget. *Ecosystems*, 10(1), 172-185.
- Colt, J. (2012). Dissolved Gas Concentration in Water: Computation as Functions of Temperature, Salinity and Pressure. *Elsevier*.
- Cramer, W., Bondeau, A., Woodward, F. I., Prentice, I. C., Betts, R. A., Brovkin, V., ... & Young-Molling, C. (2001). Global response of terrestrial ecosystem structure and function to CO₂ and climate change: results from six dynamic global vegetation models. *Global change biology*, 7(4), 357-373.
- Crump, B. C., Amaral-Zettler, L. A., & Kling, G. W. (2012). Microbial diversity in arctic freshwaters is structured by inoculation of microbes from soils. *The ISME journal*, 6(9), 1629-1639.
- Crump, B. C., Kling, G. W., Bahr, M., & Hobbie, J. E. (2003). Bacterioplankton community shifts in an arctic lake correlate with seasonal changes in organic matter source. *Applied and Environmental Microbiology*, 69(4), 2253-2268.
- Dawson, J. J. C., Soulsby, C., Tetzlaff, D., Hrachowitz, M., Dunn, S. M., & Malcolm, I. A. (2008). Influence of hydrology and seasonality on DOC exports from three contrasting upland catchments. *Biogeochemistry*, 90(1), 93-113.
- Ebel, B. A., & Moody, J. A. (2013). Rethinking infiltration in wildfire-affected soils. *Hydrological Processes*, 27(10), 1510-1514.
- Eiler, A., Langenheder, S., Bertilsson, S., & Tranvik, L. J. (2003). Heterotrophic bacterial growth efficiency and community structure at different natural organic carbon concentrations. *Applied and environmental microbiology*, 69(7), 3701-3709.
- Enright, N. J., Fontaine, J. B., Bowman, D. M., Bradstock, R. A., & Williams, R. J. (2015). Interval squeeze: altered fire regimes and demographic responses interact to threaten woody species persistence as climate changes. *Frontiers in Ecology and the Environment*, 13(5), 265-272.

- Fierer, N., Craine, J. M., McLauchlan, K., & Schimel, J. P. (2005). Litter quality and the temperature sensitivity of decomposition. *Ecology*, *86*(2), 320-326.
- Fierer, N., Morse, J. L., Berthrong, S. T., Bernhardt, E. S., & Jackson, R. B. (2007). Environmental controls on the landscape-scale biogeography of stream bacterial communities. *Ecology*, *88*(9), 2162-2173.
- Finlay, J. C. (2003). Controls of streamwater dissolved inorganic carbon dynamics in a forested watershed. *Biogeochemistry*, *62*(3), 231-252.
- Finlay, J. C., & Kendall, C. (2007). Stable isotope tracing of temporal and spatial variability in organic matter sources to freshwater ecosystems. *Stable isotopes in ecology and environmental science*, *2*, 283-333.
- Foley, J. A., Prentice, I. C., Ramankutty, N., Levis, S., Pollard, D., Sitch, S., & Haxeltine, A. (1996). An integrated biosphere model of land surface processes, terrestrial carbon balance, and vegetation dynamics. *Global biogeochemical cycles*, *10*(4), 603-628.
- Fornwalt, P. J., Huckaby, L. S., Alton, S. K., Kaufmann, M. R., Brown, P. M., & Cheng, A. S. (2016). Did the 2002 Hayman Fire, Colorado, USA, burn with uncharacteristic severity? *Fire Ecology*, *12*(3), 117-132.
- Fornwalt, P. J., & Kaufmann, M. R. (2014). Understorey plant community dynamics following a large, mixed severity wildfire in a *Pinus ponderosa*–*Pseudotsuga menziesii* forest, Colorado, USA. *Journal of Vegetation Science*, *25*(3), 805-818.
- Frossard, A., Gerull, L., Mutz, M., & Gessner, M. O. (2012). Disconnect of microbial structure and function: enzyme activities and bacterial communities in nascent stream corridors. *The ISME journal*, *6*(3), 680-691.
- Gat, J. R. (2010). *Isotope hydrology: a study of the water cycle* (Vol. 6). World Scientific.
- Gilbertson, A. (2018). Fire History Effects on Organic Matter Processing from Hillslopes to Streams. Undergraduate Thesis. Colorado College.
- Glöckner, F. O., Zaichikov, E., Belkova, N., Denissova, L., Pernthaler, J., Pernthaler, A., & Amann, R. (2000). Comparative 16S rRNA analysis of lake bacterioplankton reveals globally distributed phylogenetic clusters including an abundant group of Actinobacteria. *Applied and environmental microbiology*, *66*(11), 5053-5065.
- Godsey, S. E., Hartmann, J., & Kirchner, J. W. (2019). Catchment chemostasis revisited: Water quality responds differently to variations in weather and climate. *Hydrological Processes*, *33*(24), 3056-3069.
- Godsey, S. E., Kirchner, J. W., & Clow, D. W. (2009). Concentration–discharge relationships reflect chemostatic characteristics of US catchments. *Hydrological Processes: An International Journal*, *23*(13), 1844-1864.
- González-Pérez, J. A., González-Vila, F. J., Almendros, G., & Knicker, H. (2004). The effect of fire on soil organic matter—a review. *Environment international*, *30*(6), 855-870.
- Guarch-Ribot, A., & Butturini, A. (2016). Hydrological conditions regulate dissolved organic matter quality in an intermittent headwater stream. From drought to storm analysis. *Science of the Total Environment*, *571*, 1358-1369.

- Hagedorn, F., Saurer, M., & Blaser, P. (2004). A ^{13}C tracer study to identify the origin of dissolved organic carbon in forested mineral soils. *European Journal of Soil Science*, 55(1), 91-100.
- Hallema, D. W., Sun, G., Bladon, K. D., Norman, S. P., Caldwell, P. V., Liu, Y., & McNulty, S. G. (2017). Regional patterns of postwildfire streamflow response in the Western United States: The importance of scale-specific connectivity. *Hydrological Processes*, 31(14), 2582-2598.
- Hart, S. C., DeLuca, T. H., Newman, G. S., MacKenzie, M. D., & Boyle, S. I. (2005). Post-fire vegetative dynamics as drivers of microbial community structure and function in forest soils. *Forest Ecology and Management*, 220(1-3), 166-184.
- Hotchkiss, E. R., & Hall, R. O. (2014). High rates of daytime respiration in three streams: Use of $\delta^{18}\text{O}-\text{O}_2$ and O_2 to model diel ecosystem metabolism. *Limnology and Oceanography*, 59(3), 798-810.
- Hotchkiss, E. R., Hall Jr, R. O., Sponseller, R. A., Butman, D., Klaminder, J., Laudon, H., ... & Karlsson, J. (2015). Sources of and processes controlling CO_2 emissions change with the size of streams and rivers. *Nature Geoscience*, 8(9), 696-699.
- Hotchkiss, E. R., Sadro, S., & Hanson, P. C. (2018). Toward a more integrative perspective on carbon metabolism across lentic and lotic inland waters. *Limnology and Oceanography Letters*, 3(3), 57-63.
- Huber, U. M., Bugmann, H. K., & Reasoner, M. A. (Eds.). (2006). Global change and mountain regions: an overview of current knowledge.
- IPCC. (2018). Summary for Policymakers. In: *Global Warming of 1.5°C. An IPCC Special Report on the impacts of global warming of 1.5°C above pre-industrial levels and related global greenhouse gas emission pathways, in the context of strengthening the global response to the threat of climate change, sustainable development, and efforts to eradicate poverty.*
- Jähne, B., Münnich, K. O., Bössinger, R., Dutzi, A., Huber, W., & Libner, P. (1987). On the parameters influencing air-water gas exchange. *Journal of Geophysical Research: Oceans*, 92(C2), 1937-1949.
- Johnson, D. (2020). *Pine Gulch Fire, largest in state history, 100% contained*. KUSA.com. <https://www.9news.com/article/news/local/wildfire/pine-gulch-fire-100-contained/73-d07a2e39-73f0-43a7-af2e-9911e9d76195>.
- Jones, C. (2020). Examining the Legacy of Severe Fire on Soil Carbon Cycling in Montane Landscapes in and Around the Hayman Burn, Colorado. Undergraduate Thesis. Colorado College.
- Judd, K. E., Crump, B. C., & Kling, G. W. (2006). Variation in dissolved organic matter controls bacterial production and community composition. *Ecology*, 87(8), 2068-2079.
- Kaplan, L. A., Wiegner, T. N., Newbold, J. D., Ostrom, P. H., & Gandhi, H. (2008). Untangling the complex issue of dissolved organic carbon uptake: a stable isotope approach. *Freshwater Biology*, 53(5), 855-864.

- Khodadad, C. L., Zimmerman, A. R., Green, S. J., Uthandi, S., & Foster, J. S. (2011). Taxa-specific changes in soil microbial community composition induced by pyrogenic carbon amendments. *Soil Biology and Biochemistry*, 43(2), 385-392.
- Kirk, A. (2020). *East Troublesome Fire in Grand County now 100% contained*. KUSA.com. <https://www.9news.com/article/news/local/wildfire/east-troublesome-fire-contained/73-552f43cc-2849-469e-948d-b55fce0a01b3>.
- Knicker, H. (2007). How does fire affect the nature and stability of soil organic nitrogen and carbon? A review. *Biogeochemistry*, 85(1), 91-118.
- Langenheder, S., Bulling, M. T., Solan, M., & Prosser, J. I. (2010). Bacterial biodiversity-ecosystem functioning relations are modified by environmental complexity. *PloS one*, 5(5), e10834.
- Langenheder, S., & Lindström, E. S. (2019). Factors influencing aquatic and terrestrial bacterial community assembly. *Environmental microbiology reports*, 11(3), 306-315.
- Leibold, M. A., Holyoak, M., Mouquet, N., Amarasekare, P., Chase, J. M., Hoopes, M. F., ... & Gonzalez, A. (2004). The metacommunity concept: a framework for multi-scale community ecology. *Ecology letters*, 7(7), 601-613.
- McLauchlan, K. K., Higuera, P. E., Gavin, D. G., Perakis, S. S., Mack, M. C., Alexander, H., ... & Enders, S. K. (2014). Reconstructing disturbances and their biogeochemical consequences over multiple timescales. *BioScience*, 64(2), 105-116.
- McMurdie, P. J., & Holmes, S. (2013). phyloseq: an R package for reproducible interactive analysis and graphics of microbiome census data. *PloS one*, 8(4), e61217.
- Mei, Y., Hornberger, G. M., Kaplan, L. A., Newbold, J. D., & Aufdenkampe, A. K. (2012). Estimation of dissolved organic carbon contribution from hillslope soils to a headwater stream. *Water Resources Research*, 48(9).
- Miller, S. (2020, October 30). *'This season is off the charts': Colorado fights the worst wildfires in its recent history*. The Guardian. <https://www.theguardian.com/us-news/2020/oct/30/colorado-fires-cameron-peak-east-troublesome>.
- Moody, J. A., & Martin, D. A. (2001). Initial hydrologic and geomorphic response following a wildfire in the Colorado Front Range. *Earth Surface Processes and Landforms: The Journal of the British Geomorphological Research Group*, 26(10), 1049-1070.
- Morton, D. C., Roessing, M. E., Camp, A. E., & Tyrrell, M. L. (2003). Assessing the environmental, social, and economic impacts of wildfire. *GISF Research Paper*, 1.
- Mulholland, P. J., Fellows, C. S., Tank, J. L., Grimm, N. B., Webster, J. R., Hamilton, S. K., ... & McDowell, W. H. (2001). Inter-biome comparison of factors controlling stream metabolism. *Freshwater biology*, 46(11), 1503-1517.
- Murphy, S. F., McCleskey, R. B., Martin, D. A., Writer, J. H., & Ebel, B. A. (2018). Fire, flood, and drought: extreme climate events alter flow paths and stream chemistry. *Journal of Geophysical Research: Biogeosciences*, 123(8), 2513-2526.
- Oksanen, J., Kindt, R., Legendre, P., O'Hara, B., Stevens, M. H. H., Oksanen, M. J., & Suggests, M. A. S. S. (2007). *The vegan package*. Community Ecology Package, 10(631-637), 719.

- Olefeldt, D., Turetsky, M. R., & Blodau, C. (2013). Altered composition and microbial versus UV-mediated degradation of dissolved organic matter in boreal soils following wildfire. *Ecosystems*, *16*(8), 1396-1412.
- Öquist, M. G., Wallin, M., Seibert, J., Bishop, K., & Laudon, H. (2009). Dissolved inorganic carbon export across the soil/stream interface and its fate in a boreal headwater stream. *Environmental science & technology*, *43*(19), 7364-7369.
- Pacala, S. W., Hurtt, G. C., Baker, D., Peylin, P., Houghton, R. A., Birdsey, R. A., ... & Field, C. B. (2001). Consistent land-and atmosphere-based US carbon sink estimates. *Science*, *292*(5525), 2316-2320.
- Palmer, S. M., Hope, D., Billett, M. F., Dawson, J. J., & Bryant, C. L. (2001). Sources of organic and inorganic carbon in a headwater stream: evidence from carbon isotope studies. *Biogeochemistry*, *52*(3), 321-338.
- Prentice, I. C., Farquhar, G. D., Fasham, M. J. R., Goulden, M. L., Heimann, M., Jaramillo, V. J., ... & Wallace, D. W. (2001). *The carbon cycle and atmospheric carbon dioxide*. Cambridge University Press.
- Pressler, Y., Moore, J. C., & Cotrufo, M. F. (2019). Belowground community responses to fire: meta-analysis reveals contrasting responses of soil microorganisms and mesofauna. *Oikos*, *128*(3), 309-327.
- Raymond, P. A., & Saiers, J. E. (2010). Event controlled DOC export from forested watersheds. *Biogeochemistry*, *100*(1), 197-209.
- Reid, C. E., Brauer, M., Johnston, F. H., Jerrett, M., Balmes, J. R., & Elliott, C. T. (2016). Critical review of health impacts of wildfire smoke exposure. *Environmental health perspectives*, *124*(9), 1334-1343.
- Rhoades, C. C., Chow, A. T., Covino, T. P., Feghel, T. S., Pierson, D. N., & Rhea, A. E. (2019). The legacy of a severe wildfire on stream nitrogen and carbon in headwater catchments. *Ecosystems*, *22*(3), 643-657.
- Rhoades, C. C., Entwistle, D., & Butler, D. (2011). The influence of wildfire extent and severity on streamwater chemistry, sediment and temperature following the Hayman Fire, Colorado. *International Journal of Wildland Fire*, *20*(3), 430-442.
- Santos, F., Russell, D., & Berhe, A. A. (2016). Thermal alteration of water extractable organic matter in climate sequence soils from the Sierra Nevada, California. *Journal of Geophysical Research: Biogeosciences*, *121*(11), 2877-2885.
- Smith, H. G., Sheridan, G. J., Lane, P. N., Nyman, P., & Haydon, S. (2011). Wildfire effects on water quality in forest catchments: a review with implications for water supply. *Journal of Hydrology*, *396*(1-2), 170-192.
- Stelzer, R. S., Heffernan, J., & Likens, G. E. (2003). The influence of dissolved nutrients and particulate organic matter quality on microbial respiration and biomass in a forest stream. *Freshwater Biology*, *48*(11), 1925-1937.

- Stevens, M. R. (2013). *Analysis of postfire hydrology, water quality, and sediment transport for selected streams in areas of the 2002 Hayman and Hinman fires, Colorado (No. 2012-5267)*. US Geological Survey.
- Stevens-Rumann, C. S., Kemp, K. B., Higuera, P. E., Harvey, B. J., Rother, M. T., Donato, D. C., ... & Veblen, T. T. (2018). Evidence for declining forest resilience to wildfires under climate change. *Ecology letters*, *21*(2), 243-252.
- Strickland, M. S., Osburn, E., Lauber, C., Fierer, N., & Bradford, M. A. (2009). Litter quality is in the eye of the beholder: initial decomposition rates as a function of inoculum characteristics. *Functional Ecology*, *23*(3), 627-636.
- Striegl, R. G., Aiken, G. R., Dornblaser, M. M., Raymond, P. A., & Wickland, K. P. (2005). A decrease in discharge-normalized DOC export by the Yukon River during summer through autumn. *Geophysical Research Letters*, *32*(21).
- Thomas, F., Hehemann, J. H., Rebuffet, E., Czjzek, M., & Michel, G. (2011). Environmental and gut Bacteroidetes: the food connection. *Frontiers in microbiology*, *2*, 93.
- USFS. (2020). *Overview Summary: Cameron Peak*. https://inciweb.nwcg.gov/photos/COARF/2020-08-14-0842-Cameron-Peak/related_files/pict20201111-154527-0.pdf.
- Vannote, R. L., Minshall, G. W., Cummins, K. W., Sedell, J. R., & Cushing, C. E. (1980). The river continuum concept. *Canadian journal of fisheries and aquatic sciences*, *37*(1), 130-137.
- Vellend, M. (2010). Conceptual synthesis in community ecology. *The Quarterly Review of Biology*, *85*(2), 183-206.
- Wanninkhof, R. (1992). Relationship between wind speed and gas exchange over the ocean. *Journal of Geophysical Research: Oceans*, *97*(C5), 7373-7382.
- Weishaar, J. L., Aiken, G. R., Bergamaschi, B. A., Fram, M. S., Fujii, R., & Mopper, K. (2003). Evaluation of specific ultraviolet absorbance as an indicator of the chemical composition and reactivity of dissolved organic carbon. *Environmental science & technology*, *37*(20), 4702-4708.
- Westerling, A. L. (2016). Increasing western US forest wildfire activity: sensitivity to changes in the timing of spring. *Philosophical Transactions of the Royal Society B: Biological Sciences*, *371*(1696), 20150178.
- Westerling, A. L., Hidalgo, H. G., Cayan, D. R., & Swetnam, T. W. (2006). Warming and earlier spring increase western US forest wildfire activity. *Science*, *313*(5789), 940-943.
- Wiegner, T. N., Kaplan, L. A., Newbold, J. D., & Ostrom, P. H. (2005). Contribution of dissolved organic C to stream metabolism: a mesocosm study using ¹³C-enriched tree-tissue leachate. *Journal of the North American Benthological Society*, *24*(1), 48-67.
- Xu, N., & Saiers, J. E. (2010). Temperature and hydrologic controls on dissolved organic matter mobilization and transport within a forest topsoil. *Environmental science & technology*, *44*(14), 5423-5429.

- Yard, M. D., Bennett, G. E., Mietz, S. N., Coggins Jr, L. G., Stevens, L. E., Hueftle, S., & Blinn, D. W. (2005). Influence of topographic complexity on solar insolation estimates for the Colorado River, Grand Canyon, AZ. *Ecological Modelling*, 183(2-3), 157-172.
- Zeglin, L. H. (2015). Stream microbial diversity in response to environmental changes: review and synthesis of existing research. *Frontiers in microbiology*, 6, 454.
- Zwart, G., Crump, B. C., Kamst-van Agterveld, M. P., Hagen, F., & Han, S. K. (2002). Typical freshwater bacteria: an analysis of available 16S rRNA gene sequences from plankton of lakes and rivers. *Aquatic microbial ecology*, 28(2), 141-155.